Synthesis of naturally occurring phosphatidylinositol 3,4,5trisphosphate [PtdIns $(3,4,5)P_3$] and its diastereoisomers

Piers R. J. Gaffney and Colin B. Reese

Department of Chemistry, King's College London, Strand, London, UK WC2R 2LS

Received (in Cambridge, UK) 7th September 2000, Accepted 15th November 2000 First published as an Advance Article on the web 2nd January 2001



The chemical synthesis of naturally occurring phosphatidylinositol 3,4,5-trisphosphate [PtdIns(3,4,5) P_3] **2**, in which the 1- and 2-hydroxy functions of the glycerol moiety are esterified with stearic and arachidonic acid, respectively, is described. The synthesis of three diastereoisomers **41**, **42** and **43** of PtdIns(3,4,5) P_3 is also reported.

Introduction

The identification in 1983 by Berridge and his co-workers¹ of D-myo-inositol 1,4,5-trisphosphate $[Ins(1,4,5)P_3]$ **1** as a cellular second messenger is one of the seminal discoveries of modern biology. More recently, 1-O-(2-O-arachidonoyl-1-O-stearoyl-*sn*-glycero-3-phospho)-D-myo-inositol 3,4,5-trisphosphate [PtdIns(3,4,5)P_3] **2**, which is usually referred to as phosphatidylinositol 3,4,5-trisphosphate, has also been shown to act as a key second messenger² in cell regulation.

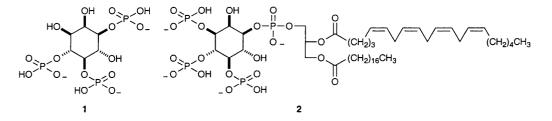
In our original synthesis of $Ins(1,4,5)P_3$ 1, we adopted a strategy³ that depended solely on the use of acid-labile and base-labile protecting groups. Indeed, unlike most subsequent workers in this field,⁴ we particularly avoided the use of benzyl and related protecting groups that are removable by hydrogenolysis. This strategy is particularly suitable for the synthesis of naturally occurring $PtdIns(3,4,5)P_3$ 2 as the olefinic double bonds in the arachidonoyl residue are clearly susceptible to catalytic hydrogenation. Prior to the preliminary publication^{5,6} of the present work, only the synthesis of $PtdIns(3,4,5)P_3$ analogues containing saturated fatty acid residues in the glycerol moiety had been reported.7-13 Following our preliminary publication,⁶ Watanabe and Nakatomi published^{14,15} an alternative synthesis of naturally occurring PtdIns $(3,4,5)P_3$ 2. We now report the full experimental details of our synthesis of PtdIns $(3,4,5)P_3$ 2 and three of its diastereoisomers.

Results and discussion

Our synthesis involved four main stages. In the first stage, the preparation of the required enantiomeric 1,2-di-O-acylglycerol derivatives was undertaken.⁵ The second stage involved the preparation of suitably protected enantiomeric *myo*-inositol 1-phosphates. In stage three, the glycerol and inositol building blocks were coupled together and the 3-, 4- and 5-hydroxy functions of the inositol residues in the coupled products were phosphorylated. In the final stage, all of the protecting groups were removed to give PtdIns(3,4,5) P_3 **2** and its diastereoisomers.

The first main stage of the synthesis is indicated in outline in Scheme 1. Enantiomerically pure 2,3-O-isopropylidene-snglycerol¹⁶ 3 was treated (Scheme 1a) with a slight excess of stearoyl chloride (R¹COCl) in the presence of triethylamine and a catalytic quantity of 4-(dimethylamino)pyridine (DMAP) in dichloromethane to give its 1-O-stearoyl derivative (step i). The product was treated with trifluoroacetic acid (TFA) in 2,2,2-trifluoroethanol-triethyl borate (1 : 8 v/v) solution at room temperature to give 1-O-stearoyl-sn-glycerol¹⁷ 4 as a crystalline solid in good isolated yield. Under these acidic conditions, acyl migration, which can lead to racemization, is almost completely avoided. Other workers¹⁸ had previously unblocked 2,3-O-isopropylidene-1-O-palmitoylglycerol with boric acid in 2-methoxyethanol at 100 °C. The 1-O-stearoyl derivative 4 was then allowed to react with a 10% excess of 9-phenylxanthen-9-ol¹⁹ (PxOH) in glacial acetic acid at room temperature to give 3-O-(9-phenylxanthen-9-yl)-1-Ostearoyl-sn-glycerol 5 as the major product. This compound 5 was treated with a $\approx 20\%$ excess of arachidonic acid (R²CO₂H) in the presence of 2,6-dichlorobenzoyl chloride²⁰ and 1-methylimidazole in dichloromethane at room temperature (Scheme 1a, step iv). Finally, the 9-phenylxanthen-9-yl protecting group²¹ was removed under very mild conditions by treatment with dichloroacetic acid in the presence of pyrrole,²² which is a very efficient carbocation scavenger, in dichloromethane solution at room temperature. In this way, 2-O-arachidonoyl-1-O-stearoylsn-glycerol 6, which is the glycerol building block required for the synthesis of naturally occurring $PtdIns(3,4,5)P_3$ 2, was obtained. The overall yield for the five steps was ≈48%. In the same way, 1,2-O-isopropylidene-sn-glycerol²³ 7 was converted in five steps (Scheme 1b and Experimental section) into 2-Oarachidonoyl-3-O-stearoyl-sn-glycerol 8.

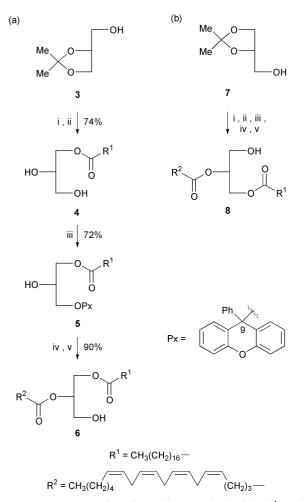
The enantiomeric purity of 2-*O*-arachidonyl-1-*O*-stearoylsn-glycerol **6** was estimated by converting it (Scheme 2) into its 3-*O*-(–)-camphanyl ester **10** and then determining the diastereoisomeric purity of this product by ¹³C NMR spectroscopy. The region of the ¹³C NMR spectrum containing the resonance signals of the glycerol carbon atoms of compound **10** is illustrated in Fig. 1a. The most downfield signal at δ 68.58 may be assigned to the resonance of *C*-2 of the glycerol moiety.



192 J. Chem. Soc., Perkin Trans. 1, 2001, 192–205

This journal is © The Royal Society of Chemistry 2001

DOI: 10.1039/b007267m



Scheme 1 Reagents and conditions: i, stearoyl chloride (R¹COCl), Et₃N, DMAP, 0 °C to room temp., 50 min; ii, CF₃CO₂H (TFA), CF₃CH₂OH, B(OEt)₃, room temp., 5 h; iii, 9-phenylxanthen-9-ol (PxOH), AcOH, room temp. to <35 °C, ≈ 15 mmHg; iv, arachidonic acid (R²COCl), 2,6-Cl₂C₆H₃COCl, 1-methylimidazole, CH₂Cl₂, room temp., 80 min; v, Cl₂CHCO₂H, pyrrole, room temp., 5 min.

The corresponding region of the ¹³C NMR spectrum of the 3-*O*-(–)-camphanyl ester of racemic 2-*O*-arachidonoyl-1-*O*-stearoylglycerol is illustrated in Fig. 1b. The signal at δ 68.73 in Fig. 1b is assigned to the C-2 resonance of the diastereo-isomeric 1-*O*-(–)-camphanyl ester **11** of 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8**. From a close examination of Fig. 1a, it may be estimated that 2-*O*-arachidonoyl-3-*O*-(–)-camphanyl-1-*O*-stearoyl-*sn*-glycerol **10** is contaminated with $\approx 2\%$ of the other diastereoisomer **11**. It follows from this that the enantiomeric excess of 2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycerol **6** must be at least 96%. In the same way, the enantiomeric excess of 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8** was estimated to be $\approx 90\%$.

The second stage of the synthesis initially involved the conversion (Scheme 3a) of *myo*-inositol **12** into 6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxy-tetrahydropyran-4-yl)-D-*myo*-inositol **21** and its enantiomer

22. *myo*-Inositol **12** was first allowed to react with *p*-anisoyl chloride in pyridine solution (step i) to give its penta-anisoyl derivative **13**, which was isolated as a pure crystalline solid in 72% yield. Under the reaction conditions, acylation of the axially disposed 2-hydroxy function occurred only to a negligible extent. Acid-catalysed reaction of compound **13** with the enol ether²⁴ **14**, followed by deacylation (steps ii and iii) gave the 2-*O*-Mthp (4-methoxytetrahydropyran-4-yl) derivative²⁴ **15** as a crystalline solid in 81% isolated yield. Reaction between compound **15** and Markiewicz reagent²⁵ **16**, in the presence of imidazole and triethylamine in hexamethyl-phosphoric triamide (HMPA) solution (step iv), gave the

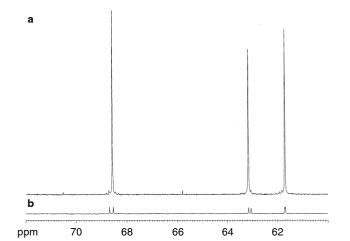
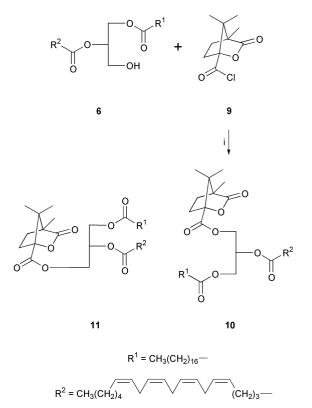
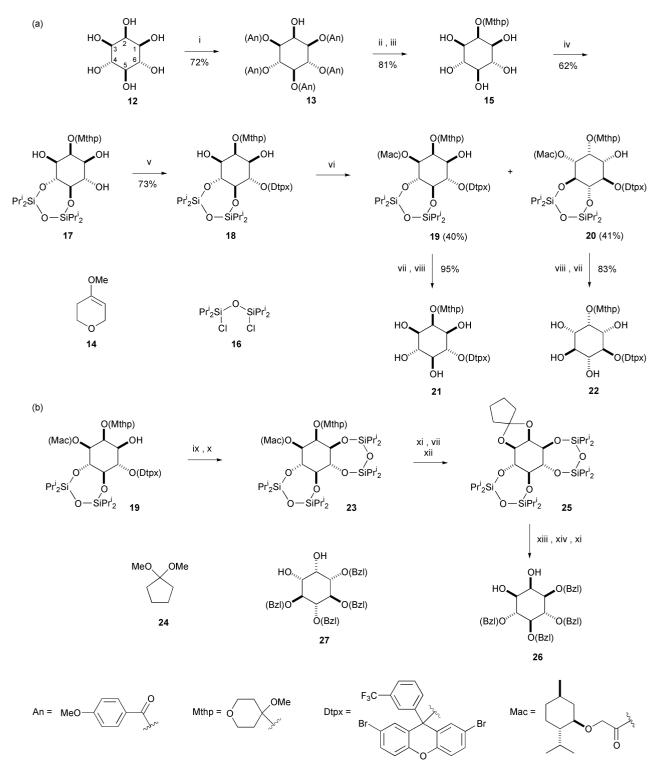


Fig. 1 Region (δ 60–70) of ¹³C NMR spectra (in CDCl₃) of (a) tris-*O*-acylglycerol derivative **10** and (b) an equimolar mixture of compound **10** and its diastereoisomer **11**.



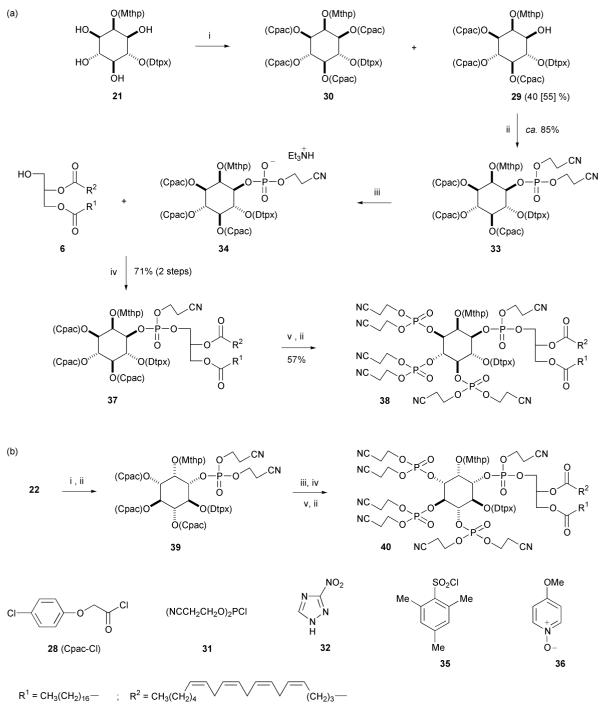
Scheme 2 Reagents and conditions: i, 1-methylimidazole, CH₂Cl₂, room temp.

4,5-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl) derivative 17, which was isolated as a crystalline solid in ≈62% yield. When the latter compound 17 was treated with 2,7-dibromo-9-chloro-9-[3-(trifluoromethyl)phenyl]xanthene²⁶ (DtpxCl) in pyridineacetonitrile solution (step v), the 6-O-Dtpx derivative 18 was obtained and isolated in 73% yield. This compound was allowed to react with (-)-menthoxyacetyl chloride (MacCl) in the presence of 1*H*-tetrazole in pyridine solution (step vi). Following fractionation of the products, each of the pure diastereoisomers 19 and 20 was isolated in a yield of $\approx 40\%$. These diastereoisomers may readily be distinguished by NMR spectroscopy: the signals at δ 80.25 and 79.74, respectively, in the ${}^{13}C$ NMR spectra (in CDCl₃) of diastereoisomers 19 and 20, may be assigned to the resonances of the carbon atom (*i.e.* C-3) directly attached to the (-)-menthoxyacetoxy residue. An examination of the two ¹³C NMR spectra revealed that neither diastereoisomer was detectably contaminated with the other. Following treatment with ethanolic methylamine (step vii)



Scheme 3 Reagents and conditions: i, AnCl, C_5H_5N , 0 °C to room temp., 18 h; ii, 14, Ph₃P·HBr, room temp., 40 h; iii, NaOMe, MeOH, THF, reflux, 30 min; iv, 16, imidazole, Et₃N, HMPA, room temp., 60 h; v, DtpxCl, C_5H_5N , MeCN, room temp., 80 min; vi, MacCl, 1*H*-tetrazole, DMAP, MeCN, C_5H_5N , room temp., 30 min; vii, MeNH₂, EtOH, room temp., 90 min; viii, Et₄NF, MeCN, room temp., 30 min; ix, ClCH₂CO₂H, pyrrole, CH₂Cl₂, room temp., 15 min; x, 16, imidazole, DMF, 80 °C, 7 h; xi, TsOH·H₂O, CH₂Cl₂-MeOH (1:1 v/v), room temp., 2 h; xii, 24, TsOH·H₂O, room temp., 1 h; xiii, Et₄NF, Et₄NOH, MeCN, aq. EtOH, reflux, 16 h; xiv, PhCH₂Cl, NaH, DMF, room temp., 2.5 h.

and tetraethylammonium fluoride (TEAF) (step viii), diastereoisomer 19 was converted into $6-O-\{2,7-dibromo-9-[3-(trifluoro$ $methyl)phenyl]xanthen-9-yl\}-2-<math>O-(4$ -methoxytetrahydropyran-4-yl)-D-myo-inositol 21 in 95% yield. An important piece of evidence in support of the enantiomeric purity of compound 21 is that its precursor 19 was diastereoisomerically almost pure. It was not possible to obtain satisfactory optical rotation data for compound 21. However, the circular dichroism (CD) spectrum of compound 21 and that of its enantiomer 22, which was prepared in the same way (Scheme 3a) from the L-inositol derivative 20, are virtually exact mirror images of each other. Although it seemed likely that the enantiomeric excesses of compounds **21** and **22** were both very high, it was by no means clear which enantiomer was which. This was established (Scheme 3b) by relating them to the known 1,4,5,6-tetra-*O*-benzyl-D- and -L-*myo*-inositol²⁷ **26** and **27**. The (-)-menthoxy-acetyl derivative **19** was treated first with chloroacetic acid in the presence of pyrrole in dichloromethane solution (Scheme 3b, step ix) and then with Markiewicz reagent **16** under rather forcing conditions (step x) to give the bis(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl) derivative **23**. After the Mthp and Mac groups had been removed by treatment with



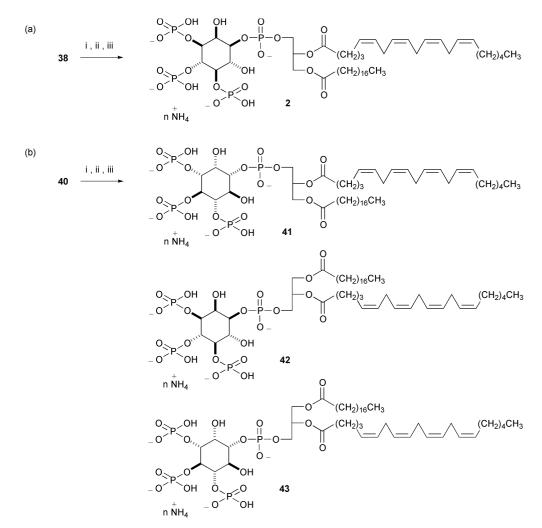
Scheme 4 Reagents and conditions: i, CpacCl 28, 1*H*-tetrazole, DMAP, MeCN, C_5H_5N , room temp., 95 min; ii, (a) 31, 32, C_5H_5N , MeCN, room temp., 1 h; (b) 70% *t*-BuO₂H, room temp., 90 min; iii, Et₃N, MeCN, room temp., 16 h; iv, 35, 36, MeCN, C_5H_5N , room temp. 1 h; v, N_2H_4 ·H₂O, MeCN, room temp., 90 min.

toluene-4-sulfonic acid (PTSA) in methanol–dichloromethane and ethanolic methylamine, respectively, the 2,3-diol system was protected (as a cyclopentylidene acetal²⁸) by treatment with 1,1-dimethoxycyclopentane **24** in the presence of PTSA (step xii) to give compound **25**. Following the removal of the two 1,1,3,3-tetraisopropyldisiloxane-1,3-diyl groups (step xiii), the 1-, 4-, 5- and 6-hydroxy functions were benzylated by treatment with benzyl chloride and sodium hydride in dimethylformamide (DMF) (step xiv). Finally, the cyclopentylidene protecting group was removed by acid-catalyzed methanolysis to give 1,4,5,6-tetra-*O*-benzyl-D-*myo*-inositol **26**. The identity of enantiomer **26** was clear from its specific rotation ($[a]_{D}^{20} + 22.7 \dagger$), which was close to the value reported²⁷ in the literature. In the same way (Scheme 3b), diastereoisomer **20**, the precursor of enantiomer **22**, was converted into 1,4,5,6-tetra-*O*-benzyl-L-*myo*-inositol **27** ($[a]_{D}^{20} - 21.7$).

The rest of the second stage of the synthesis of naturally occurring $PtdIns(3,4,5)P_3 2$ (*i.e.*, the conversion of 6-O-Dtpx-2-O-Mthp-D-myo-inositol 21 into the protected D-myo-inositol 1-phosphate building block 34) and the third stage (*i.e.*, the coupling reaction and the phosphorylation of the 3-, 4- and 5-hydroxy functions of the inositol moiety in the coupled product) are indicated in outline in Scheme 4a.

Reaction between 6-O-Dtpx-2-O-Mthp-D-*myo*-inositol **21** and (4-chlorophenoxy)acetyl chloride (CpacCl) **28** in the presence both of DMAP and 1*H*-tetrazole in acetonitrile–pyridine solution gave a mixture of the 3,4,5-tris-O-(4-chloro-

[†] Throughout this paper, $[a]_D$ -values are given in units of $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$.



Scheme 5 Reagents and conditions: i, $(Me_2N)_2C=NH$ (TMG), Me_3SiCl , MeCN, room temp., 16 h; ii, NH_3 , aq. MeOH, room temp.; iii, (a) AcOH-water (2:1 v/v), room temp., 90 min; (b) aq. NH₃, MeOH, room temp.

phenoxy)acetyl and 1,3,4,5-tetrakis-O-(4-chlorophenoxy)acetyl derivatives²⁹ 29 and 30. Following fractionation of the products, the desired tris[(4-chlorophenoxy)acetate] 29 was isolated in 40% yield. However, the starting material 21 could be regenerated from the tetrakis[(4-chlorophenoxy)acetate] 30 by treating it with alcoholic methylamine. Thus the effective yield of the desired tris[(4-chlorophenoxy)acetate] 29 was 55%, based on the 6-O-Dtpx-2-O-Mthp-D-myo-inositol 21 consumed. Phosphorylation of the tris[(4-chlorophenoxy)acetate] 29 with di(2-cyanoethyl) phosphorochloridite³⁰ 31 (see Experimental section) in the presence of 3-nitro-1,2,4-1H-triazole 32 in pyridine-acetonitrile, followed by oxidation of the resulting phosphite triester with tert-butyl hydroperoxide³¹ (Scheme 4, step ii) gave the 1-[di(2-cyanoethyl) phosphate] 33 in ${\approx}85\%$ isolated yield. This material was treated with triethylamine in acetonitrile (step iii) to give the triethylammonium salt of the corresponding 1-[(2-cyanoethyl) phosphate] 34.

In the third step of the synthesis, the triethylammonium salt **34** was allowed to react with 2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycerol **6** and mesitylene-2-sulfonyl chloride **35** in the presence of 4-methoxypyridine 1-oxide ³² **36** (Scheme 4a, step iv) to give the fully protected coupled product **37** in 71% overall yield for steps iii and iv. In order to complete this stage of the synthesis, the (4-chlorophenoxy)acetyl groups were removed from the coupled product **37** by hydrazinolysis (step v). Under the reaction conditions, both the stearoyl and arachidonoyl residues remained completely intact. The released 3-, 4- and 5-hydroxy functions were then phosphorylated by treatment first with di(2-cyanoethyl) phosphorochloridite **31**, followed by oxidation of the intermediate trisphosphite with *tert*-butyl

hydroperoxide to give the fully protected $PtdIns(3,4,5)P_3$ **38**. This material was isolated from the products in $\approx 57\%$ overall yield for the three steps (v, iia and iib), starting from the tris[(4-chlorophenoxy)acetyl] derivative **37**.

In the same way, 6-O-Dtpx-2-O-Mthp-L-myo-inositol 22 was converted into the L-enantiomer of its tris[(4-chlorophenoxy)acetate] (*i.e.*, the enantiomer of compound **29**), which in turn was converted (Scheme 4, step ii) into the corresponding 1-[di(2-cyanoethyl) phosphate] 39. Following treatment with triethylamine in acetonitrile (step iii), the product (i.e., the enantiomer of triethylammonium salt 34) was then coupled with 2-O-arachidonoyl-1-O-stearoyl-sn-glycerol 6. Following the same three-step procedure (steps v, iia and iib), the initial coupling product, which corresponded to the tris[(4-chlorophenoxy)acetyl] derivative 37, was converted into the L-myoinositol analogue 40 of the fully protected $PtdIns(3,4,5)P_3$ 38. The yields obtained in the various steps are indicated in the Experimental section. Two additional diastereoisomers of the fully protected PtdIns $(3,4,5)P_3$ **38** were prepared by first coupling both intermediate phosphodiesters (i.e., 34 and its enantiomer) with 2-O-arachidonoyl-3-O-stearoyl-sn-glycerol 8 and then following the same three-step procedure (Scheme 4, steps v, iia and iib). The preparative details and the yields obtained are again indicated in the Experimental section.

The three-step procedure followed in the unblocking of the fully protected PtdIns $(3,4,5)P_3$ **38** is indicated in outline in Scheme 5. Arguably the most critical step was the removal of the seven 2-cyanoethyl protecting groups. In order to ensure that the 3-, 4- and 5-phosphate residues were completely unblocked, a modification of Evans' procedure,³³ in which

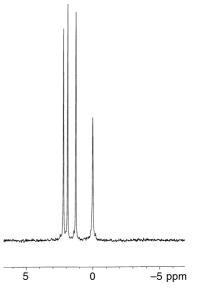


Fig. 2 31 P NMR spectrum (in CD₃OD–D₂O) of the ammonium salt of PtdIns(3,4,5) P_3 2.

chlorotrimethylsilane was added to a solution of fully protected PtdIns(3,4,5) P_3 **38** and N^1, N^1, N^3, N^3 -tetramethylguanidine (TMG) in acetonitrile solution, was adopted. After subsequent treatment with aq. methanolic ammonia (step ii), the products were kept in acetic acid–water solution (step iii) in order to remove the acid-labile 2-*O*-Mthp and 6-*O*-Dtpx groups. The ammonium salt of PtdIns(3,4,5) P_3 **2** was then isolated as an off-white hygroscopic solid in ≈96% yield. The material was characterized on the basis of ¹H and ³¹P NMR and MALDI-TOF mass spectroscopic data; as can be seen from Fig. 2, its ³¹P NMR spectrum displays four well resolved resonance signals. Other diastereoisomers of fully protected PtdIns(3,4,5) P_3 **38** (*i.e.*, **40** and the two diastereoisomers derived from 2-*O*arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8**) were unblocked by the same three-step procedure (Scheme 5 and Experimental section) to give products **41**, **42** and **43**.

The biological activity of this synthetic natural PtdIns- $(3,4,5)P_3$ **2** has been clearly demonstrated in a number of biological studies.³⁴⁻³⁷ Alessi *et al.*³⁴ showed that, at very low concentration, natural PtdIns $(3,4,5)P_3$ **2** was much more effective in activating PtdIns $(3,4,5)P_3$ -dependent protein kinase 1 than is the corresponding lipid in which both the arachidonoyl and stearoyl residues were replaced by saturated (*i.e.* palmitoyl) residues. These workers³⁴ further reported that the two diastereoisomers **41** and **43** derived from L-*myo*-inositol were inactive. These conclusions were essentially confirmed by Stokoe, Stephens *et al.*³⁶ in corresponding studies.

Experimental

Mps were measured with a Büchi melting-point apparatus and are uncorrected. ¹H NMR spectra were measured at 360 MHz with a Bruker AM 360 spectrometer. ¹³C NMR spectra were measured at 90.6 MHz with the same spectrometer. Chemical shifts are given in ppm relative to tetramethylsilane, and *J*-values are given in Hz; ³¹P NMR spectra were measured at 145.8 MHz with the same spectrometer with 85% orthophosphoric acid being used as external standard. Mass spectra were obtained using Kratos MS890 and Voyager MALDI-TOF spectrometers. Optical rotations were measured with a Perkin-Elmer 141 polarmeter. Merck silica gel 60 F₂₅₄ plates (Art 5715 and 5642) were used for TLC and were developed in the solvent systems indicated below. Merck silica gel 60 (Art 7729 and 9385) was used for short-column chromatography and Merck silanized silica gel (Art 7719) was used for reversedphase column chromatography. Acetonitrile, pyridine and triethylamine were dried by heating with calcium hydride, under reflux, for 3-5 h and were then distilled; 1-methylimidazole, TMG, DMF and HMPA were dried by heating with calcium hydride at ≈100 °C and were then distilled under reduced pressure; methanol was dried by heating under reflux with magnesium methoxide (generated in situ from magnesium turnings and methanol) and was then distilled; diethyl ether was dried over sodium wire and was then distilled; THF was dried by heating, under reflux, over sodium-potassium alloy and benzophenone and was then distilled; dichloromethane was dried over phosphorus pentaoxide and was then distilled; toluene was dried by heating, under reflux, with sodium metal and was then distilled. All solvents were stored over 4 Å molecular sieves in sealed containers. Petroleum spirit refers to the fraction with distillation range 60-80 °C, unless stated otherwise.

1-O-Stearoyl-sn-glycerol 4; $R^1 = C_{17}H_{35}$

Triethylamine (4.15 cm³, 29.8 mmol), followed by stearoyl chloride (6.03 cm³, 17.9 mmol), was added to a rapidly stirred solution of 2,3-O-isopropylidene-sn-glycerol 3 (1.96 g, 14.8 mmol) and DMAP (0.182 g, 1.5 mmol) in dry dichloromethane (40 cm³) at 0 °C (ice–water-bath). After 5 min, the reactants were allowed to warm to room temperature. After a further period of 45 min, water (0.64 cm³) was added and the products were concentrated under reduced pressure. The residue was stirred with hexane (150 cm³) and the insoluble material was removed by filtration. Silanized silica gel (50 g) was added to the filtrate and the resulting mixture was evaporated under reduced pressure. Acetonitrile-water (1:1 v/v; 600 cm³) was added to the residue which was then filtered through a large sintered glass funnel. The filtrate was discarded. The residue was then washed with acetonitrile (600 cm³) and, after toluene (200 cm³) had been added, the washings were concentrated under reduced pressure. The crude 1-O-stearoyl-2,3-O-isopropylidene-sn-glycerol (6.09 g) thereby obtained was dissolved in triethyl borate (80 cm³) at room temperature, and 2,2,2-trifluoroethanol (10 cm³) and TFA (10 cm³) were added. The reactants were stirred at room temperature. After 5 h, the products were evaporated under reduced pressure. Dry toluene (50 cm³) was added and the resulting solution was reevaporated under reduced pressure. The process was repeated and the residue was dissolved in methanol (200 cm³). Water (200 cm³; the first 20 cm³ dropwise) was added. The colourless solid precipitate was collected by filtration, washed with water and then dissolved in diethyl ether (600 cm³). The dried (MgSO₄) solution was evaporated under reduced pressure and the residue was crystallized twice from hexane to give the title compound (3.955 g, 74%) (Found: C, 69.7; H, 12.0. C₂₁H₄₂O₄· 0.2 H₂O requires C, 69.45; H 11.80%), mp 67.5–68.5 °C (lit.,¹⁸ 72.5–73 °C); $[a]_{D}^{20}$ +3.66 (c 4, C₅H₅N) {lit., ¹⁸ $[a]_{D}^{25}$ +3.55 (c 5.24, C₅H₅N)}; δ_H (CDCl₃) 0.87 (3 H, t, J 6.8), 1.25 (28 H, m), 1.60 (2 H, m), 2.34 (2 H, t, J 7.6), 2.60 (1 H, br), 2.97 (1 H, br), 3.57 (1 H, dd, J 5.9 and 11.5), 3.69 (1 H, dd, J 3.7 and 11.5), 3.92 (1 H, m), 4.16 (2 H, m); $\delta_{\rm C}$ (CDCl₃) 14.19, 22.76, 24.97, 29.20, 29.33, 29.43, 29.53, 29.68, 29.73, 29.77, 31.99, 34.23, 63.42, 65.18, 70.33, 174.49.

3-O-Stearoyl-sn-glycerol

1,2-*O*-Isopropylidene-*sn*-glycerol 7 (1.00 g, 7.56 mmol) was converted into 3-*O*-stearoyl-*sn*-glycerol (1.49 g, 55%) (Found: C, 69.9; H, 11.9. $C_{21}H_{42}O_4$ requires C, 70.35; H, 11.81%) by the same two-step procedure (see the above preparation of 1-*O*-stearoyl-*sn*-glycerol); it had mp 68.5 °C; $[a]_{19}^{19}$ -3.41 (*c* 3.99, C_5H_5N). The ¹H and ¹³C NMR spectra of 3-*O*-stearoyl-*sn*-glycerol were identical with the corresponding spectra (see above) of the 1-*O*-stearoyl enantiomer **4**.

3-*O*-(9-Phenylxanthen-9-yl)-1-*O*-stearoyl-*sn*-glycerol 5; $R^1 = C_{17}H_{35}$

A solution of 1-O-stearoyl-sn-glycerol 4 (2.87 g, 8.0 mmol) and PxOH (2.42 g, 8.8 mmol) in glacial acetic acid (100 cm³) was evaporated under reduced pressure (bath temperature <35 °C). The residue was re-dissolved in glacial acetic acid $(2 \times 50 \text{ cm}^3)$ and the resulting solution was re-evaporated. A solution of the residue in hexane (100 cm³) was washed with saturated aq. sodium hydrogen carbonate $(2 \times 50 \text{ cm}^3)$. The dried (MgSO₄) organic layer was evaporated under reduced pressure. The residue was stirred with hexane (30 cm^3) and the insoluble material was removed by filtration. The filtrate was applied to a column of silica gel (100 g). The column was then eluted with hexaneethyl acetate (100:0 to 74:26 v/v) containing $\approx 0.1\%$ pyridine. Early fractions, which were eluted with hexane-ethyl acetate (96:4 to 94:6 v/v), were evaporated under reduced pressure to give putative 2,3-bis-O-(9-phenylxanthen-9-yl)-1-O-stearoyl-snglycerol (1.05 g); $R_f 0.75$ [hexane-diethyl ether (1:1 v/v)]. Later fractions, which were eluted with hexane-ethyl acetate (86:14 to 77:23 v/v), were combined, washed successively with 0.2 mol dm⁻³ sodium acetate buffer (pH 5) and saturated aq. sodium hydrogen carbonate, dried (MgSO₄) and evaporated under reduced pressure to give the title compound as a colourless oil (3.58 g, $\approx 72\%$); $R_{\rm f}$ 0.48 [hexane-diethyl ether (1:1 v/v)]; $[a]_{\rm D}^{20}$ +4.1 (c 2.02, toluene); $\delta_{\rm H}$ (CDCl₃) 0.80 (3 H, m), 1.18 (28 H, m), 1.46 (2 H, m), 2.15 (2 H, t, J 7.6), 2.30 (1 H, br), 2.94 (2 H, m), 3.85 (1 H, m), 4.07 (2 H, m), 6.94 (2 H, m), 7.0-7.3 (11 H, m); $\delta_{\rm C}$ (CDCl₃) 14.57, 23.13, 25.26, 29.56, 29.68, 29.80, 29.90, 30.05, 30.10, 30.14, 32.36, 34.55, 64.17, 65.92, 69.49, 76.11, 116.80, 123.04, 124.05, 126.85, 127.16, 128.31, 129.74, 129.93, 148.88, 151.69, 174.39.

1-O-(9-Phenylxanthen-9-yl)-3-O-stearoyl-sn-glycerol

3-*O*-Stearoyl-*sn*-glycerol (1.401 g, 3.9 mmol) was converted into 1-*O*-(9-phenylxanthen-9-yl)-3-*O*-stearoyl-*sn*-glycerol (1.84 g, \approx 76%) by the same procedure [see the above preparation of 3-*O*-(9-phenylxanthen-9-yl)-1-*O*-stearoyl-*sn*-glycerol **5**; R¹ = C₁₇H₃₅)]; it has [a]₂₀²⁰ -4.1 (*c* 3.03, toluene). The ¹H and ¹³C NMR spectra of 1-*O*-(9-phenylxanthen-9-yl)-3-*O*-stearoyl-*sn*-glycerol were virtually identical with the corresponding spectra of the above enantiomer.

2-*O*-Arachidonoyl-1-*O*-stearoyl-*sn*-glycerol 6; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$

1-Methylimidazole (1.38 cm³, 17.3 mmol), a solution of arachidonic acid (1.053 g, 3.46 mmol) in dichloromethane (5 cm³), and 2,6-dichlorobenzovl chloride (0.99 cm³, 6.9 mmol) were added in turn to a stirred solution of 3-O-(9-phenylxanthen-9yl)-1-*O*-stearoyl-*sn*-glycerol **5**; $R^1 = C_{17}H_{35}$ (1.774 g, ≈ 2.9 mmol) in dichloromethane (20 cm³) at room temperature. A heavy, colourless precipitate was obtained after 2 min. After 80 min, water (0.21 cm³, 11.7 mmol) and then triethylamine (1.20 cm³, 8.6 mmol) were added. The products were evaporated under reduced pressure and the residue was triturated with diethyl ether-hexane (1:5 v/v; 3×30 cm³). The insoluble material was removed by decantation and the supernatant was washed with saturated aq. sodium hydrogen carbonate (100 cm³). The aqueous layer was back-extracted with hexane (50 cm³). The combined organic layers were washed first with 0.2 mol dm⁻³ sodium acetate buffer (pH \approx 5; 2 × 100 cm³) and then with an aqueous solution of sodium chloride and sodium hydrogen carbonate (50 cm³). The dried (MgSO₄) organic layer was evaporated under reduced pressure. A solution of the residue in hexane was applied to a column of silica gel (100 g), which was then eluted with hexane-diethyl ether (98:2 v/v). The appropriate fractions of the eluate were evaporated under reduced pressure to give an oily residue (2.57 g); $R_{\rm f}$ 0.43 [hexane-diethyl ether (9:1 v/v)]. This material was dissolved in dry dichloromethane (45 cm³), and pyrrole (1.95 cm³, 28 mmol) and dichloroacetic acid (0.47 cm³, 5.7 mmol) were added to the stirred solution at room temperature. After 5 min, the reaction solution was shaken with 0.2 mol dm⁻³ sodium acetate buffer (pH \approx 5; 150 cm³). The layers were separated and the aqueous layer was extracted with dichloromethane (50 cm³). The combined organic layers were dried (MgSO₄) and evaporated under reduced pressure, ensuring that all of the excess of pyrrole was removed. The residue was triturated with hexane-acetic acid $(99.95:0.05 \text{ v/v}; 3 \times 10 \text{ cm}^3)$. The supernatant was applied to a column of silica gel (100 g) which was then eluted with hexane–diethyl ether (100:0 to 40:60 v/v) containing $\approx 0.05\%$ acetic acid. The appropriate fractions, which were eluted with hexane-diethyl ether (80:20 to 45:55 v/v), were combined, and evaporated under reduced pressure to give the *title compound* (1.68 g, $\approx 90\%$) as an oily residue (HRMS Found: M⁺, 644.5360. $^{12}C_{41}^{1}H_{72}^{15}O_5$ requires *M*, 644.53795); $R_f 0.43$ [diethyl etherhexane (1:1 v/v)]; $[a]_{D}^{22} - 0.4$ (c 3.42, toluene); δ_{H} (CDCl₃) 0.88 (6 H, m), 1.25–1.45 (34 H, m), 1.61 (2 H, m), 1.72 (2 H, m), 2.06 (2 H, dd, J 6.9 and 13.8), 2.13 (2 H, dd, J 7.2 and 13.6), 2.34 (4 H, dt, J 7.6 and 16.6), 2.82 (6 H, m), 3.72 (2 H, d, J 5.1), 4.22 (1 H, dd, J 5.8 and 11.9), 4.33 (1 H, dd, J 4.3 and 12.0), 5.09 (1 H, m), 5.37 (8 H, m); δ_C (CDCl₃) 14.33, 14.37, 22.83, 22.95, 24.97, 25.12, 25.85, 26.71, 27.46, 29.37, 29.52, 29.57, 29.62, 29.73, 29.88, 29.92, 29.95, 31.76, 32.18, 33.86, 34.32, 61.58, 62.33, 72.41, 127.75, 128.05, 128.30, 128.51, 128.82, 128.99, 129.23, 130.71, 173.46, 174.08.

2-*O*-Arachidonoyl-3-*O*-stearoyl-*sn*-glycerol 8; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$

1-*O*-(9-Phenylxanthen-9-yl)-3-*O*-stearoyl-*sn*-glycerol (1.255 g, \approx 2.04 mmol) was allowed to react with arachidonic acid (0.746 g, 2.45 mmol), 2,6-dichlorobenzoyl chloride (0.70 cm³, 4.9 mmol) and 1-methylimidazole (0.98 cm³, 12.1 mmol) in dichloromethane (12 cm³) solution, as in the above preparation of 2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycerol, to give its 2-*O*-arachidonoyl derivative (1.794 g). This material (0.853 g) was treated as above with pyrrole (0.65 cm³, 9.4 mmol) and dichloroacetic acid (0.156 cm³, 3.2 mmol) in dichloromethane (15 cm³) to give the *title compound* (0.578 g, \approx 92% overall yield for the two steps). The ¹H and ¹³C NMR spectra of 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol were closely similar to the corresponding spectra of 2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycerol **6**; R¹ = C₁₇H₂₅, R² = C₁₉H₃₁.

Determination of the diastereoisomeric excesses of 2-*O*arachidonoyl-1(and 3)-*O*-stearoyl-*sn*-glycerol 6 and 8; $R^1 = C_{17}H_{35}, R^2 = C_{19}H_{31}$

(a) 1-Methylimidazole (0.121 cm³, 1.5 mmol) and then (-)camphanyl chloride 9 (0.111 g, 0.5 mmol) were added to a stirred solution of 2-O-arachidonoyl-1-O-stearoyl-sn-glycerol **6** (0.156 g, 0.24 mmol) in dry dichloromethane (3 cm³) at room temperature. After 45 min, hexane (50 cm³) was added and the resulting solution was washed with saturated aq. sodium hydrogen carbonate $(2 \times 50 \text{ cm}^3)$. The organic layer was separated and washed first with 0.2 mol dm⁻³ sodium acetate buffer (pH \approx 5; 50 cm³) and then with an aqueous solution of sodium acetate and sodium chloride (50 cm³). The dried (MgSO₄) organic layer was evaporated under reduced pressure and the residue was fractionated by chromatography on silica gel (10 g). The column was eluted with hexane-diethyl ether (100:0 to 76:24 v/v); the appropriate fractions, which were eluted with hexane-diethyl ether (90:10 to 80:20 v/v) were combined, and evaporated under reduced pressure to give 2-Oarachidonoyl-3-O-(-)-camphanyl-1-O-stearoyl-sn-glycerol 10; $R^{1} = C_{17}H_{35}$, $R^{2} = C_{19}H_{31}$ (0.184 g, $\approx 92\%$), R_{f} 0.44 [hexanediethyl ether (1:1 v/v)]; $\delta_{\rm H}$ (CDCl₃) 0.83 (6 H, m), 0.92 (3 H, s), 1.02 (3 H, s), 1.07 (3 H, s), 1.25 (34 H, m), 1.61 (5 H, m), 1.87 (1 H, m), 2.02 (5 H, m), 2.33 (5 H, m), 2.78 (6 H, m), 4.13 (1 H, dd, J 5.8 and 11.9), 4.27 (2 H, m), 4.45 (1 H, dd, J 4.0 and 11.9), 5.33 (9 H, m); δ_c (CDCl₃) 9.64, 14.04, 14.09, 16.61, 16.67, 22.53, 22.65, 24.62, 24.79, 25.56, 25.57, 25.59, 26.41, 27.17, 28.85, 29.06, 29.23, 29.25, 29.28, 29.33, 29.43, 29.58, 29.62, 29.66, 30.64, 31.47, 31.89, 33.43, 33.93, 54.13, 54.69, 61.71, 63.16, 68.58, 90.83, 127.76, 128.00, 128.22, 128.52, 128.64, 128.98, 130.39, 166.99, 172.43, 173.07, 177.71. An expansion of this ¹³C NMR spectrum revealed (Fig. 1a) two signals in the region of δ 68 [68.58 (98%) and 68.72 (2%)], assigned to the C-2 resonances of 2-O-arachidonoyl-3-O-(-)-camphanyl-1-Ostearoyl-sn-glycerol 10; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$ and 2-Oarachidonoyl-1-O-(-)-camphanyl-3-O-stearoyl-sn-glycerol 11; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$, respectively. The enantiomeric excess of the 1-O-stearoyl isomer was thereby estimated to be at least 96%.

(b) In the same way, 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8**; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$, was converted into its 1-*O*-(-)-camphanyl derivative **11**. Examination of the ¹³C NMR spectrum (CDCl₃) of this product revealed resonance signals at δ 68.68 (5%) and 68.83 (95%), respectively. The enantiomeric excess of 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8** was thereby estimated to be at least 90%. The ¹³C NMR spectrum (CDCl₃) of a 1:1 mixture of the two diasterioisomers (Fig. 1b), prepared by the action of (-)-camphanyl chloride **9** on the racemic modification of 2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycerol, revealed two resonance signals of equal intensity in the region of δ 68.

1,3,4,5,6-Penta-O-(p-anisoyl)-myo-inositol 13

myo-Inositol 12 (19.61 g, 0.109 mol) was added to a stirred solution of p-anisoyl chloride (96.32 g, 0.565 mol) in dry pyridine (380 cm³) at 0 °C (ice-water-bath). After 1 h, the reactants were allowed to warm to room temperature. After a further period of 18 h, water (20 cm³) was added and the products were evaporated under reduced pressure. A solution of the residue in chloroform (2 dm³) was washed first with saturated aq. sodium hydrogen carbonate (700 + 300 cm^3) and then with 4 mol dm⁻³ hydrochloric acid (350 cm³). The dried (MgSO₄) organic layer was evaporated under reduced pressure and re-dissolved in boiling dichloromethane ($\approx 2.5 \text{ dm}^3$). Petroleum spirit ($\approx 1 \text{ dm}^3$) was added until the solution became slightly turbid. After the solution had been kept at room temperature for 14 h, the title compound was collected by filtration as a colourless crystalline precipitate (66.72 g, 72%) (Found, in material recrystallized from hexane-ethyl acetate: C, 64.7; H, 4.8. C₄₆-H₄₂O₁₆ requires C, 64.94; H 4.98%), mp 234–236 °C; R_f 0.55 [chloroform–methanol (95:5 v/v)]; $\delta_{\rm H}$ (CDCl₃ + CD₂Cl₂) 3.67, 3.68, 3.72 (15 H, 3 s), 4.71 (1 H, t, J 2.2), 5.45 (2 H, dd, J 2.4 and 10.4), 5.80 (1 H, t, J 9.9), 6.21 (2 H, t, J 10.2), 6.68 (6 H, m), 6.75 (4 H, m), 7.73 (6 H, m), 7.85 (4 H, m); $\delta_{\rm C}$ (CDCl₃ + CD₂Cl₂) 55.21, 55.26, 68.62, 69.63, 70.81, 71.65, 113.38, 113.56, 121.17, 121.35, 131.59, 131.64, 131.79, 163.36, 163.63, 165.03, 165.10.

2-O-(4-Methoxytetrahydropyran-4-yl)-myo-inositol 15

Triphenylphosphine hydrobromide³⁸ (1.60 g, 4.7 mmol) was added to a stirred suspension of 1,3,4,5,6-penta-*O*-(*p*-anisoyl)*myo*-inositol **13** (39.58 g, 46.5 mmol) and 4-methoxy-3,6dihydro-2*H*-pyran²⁴ **14** (\approx 85%; 62.4 g, \approx 0.465 mol) in dichloromethane (460 cm³) at room temperature. After 40 h, triethylamine (5.0 cm³) was added and the products were concentrated under reduced pressure (water pump, bath temperature <25 °C). Diethyl ether was added to the yellow oily residue until the precipitation of a colourless solid commenced. After 5 min, additional diethyl ether (\approx 500 cm³) followed by petroleum spirit (30–40 °C; \approx 350 cm³) were then added to complete the precipitation process. The precipitate was collected by filtration t and dissolved in boiling ethyl acetatetriethylamine (99.9:0.1 v/v). Activated charcoal (≈5 g) was added to the boiling solution which, after 5 min, was filtered. Petroleum spirit was added to the hot filtrate until it became slightly turbid. After 18 h at room temperature, the colourless crystalline precipitate (44.73 g) was collected by filtration; mp 219–221 °C; R_f 0.43 [chloroform–methanol (98:2 v/v)]. This material (73.32 g, prepared in two batches) was dissolved in methanol-THF (1:4 v/v; 1.2 dm³) and the stirred solution was heated under gentle reflux. Methanolic sodium methoxide (25% w/w; 8.8 cm³, ≈38 mmol) was added with continued heating and stirring. After 30 min, the products were allowed to cool. The resulting colourless precipitate was collected by filtration, washed with diethyl ether and then dissolved in the minimum quantity of water (≈20 cm³). Aq. triethylammonium hydrogen carbonate (2 mol dm⁻³; 4 cm³) was added and the resulting solution was evaporated under reduced pressure. Crystallization of the residue from methanol-water gave the title compound (18.27 g, 81% overall yield) (Found: C, 48.85; H, 7.5. C₁₂H₂₂O₈ requires C, 48.97; H, 7.53%), mp 187 °C; $R_{\rm f}$ 0.21 [chloroform–methanol (7:3 v/v)]; $\delta_{\rm H}$ [(CD₃)₂SO–D₂O] 1.75 (4 H, m), 2.95 (1 H, t, J 8.9), 3.16 (5 H, m), 3.38 (4 H, m), 3.67 (2 H, m), 4.01 (1 H, br s); $\delta_{\rm C}$ [(CD₃)₂SO] 34.63, 48.49, 64.42, 71.05, 73.03, 73.46, 75.49, 93.36.

2-*O*-(4-Methoxytetrahydropyran-4-yl)-4,5-*O*-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-*myo*-inositol 17

Triethylamine (94 cm³, 0.67 mol) and 1,3-dichloro-1,1,3,3tetraisopropyldisiloxane²⁵ 16 (26.83 cm³, 84 mmol) were added to a mechanically stirred mixture of finely powered 2-O-(4methoxytetrahydropyran-4-yl)-myo-inositol 15 (20.57 g, 69.9 mmol), imidazole (22.84 g, 0.335 mol) and HMPA (350 cm³) at room temperature. After 60 h, the products were partitioned between ethyl acetate (1.5 dm³) and half-saturated aq. sodium hydrogen carbonate (1.0 dm³). The separated organic phase was washed with half-saturated aq. sodium hydrogen carbonate (0.5 dm^3) and the combined aqueous layers were back-extracted with ethyl acetate (200 cm³). The combined organic layers were washed successively with water $(3 \times 1.0 \text{ dm}^3)$ and saturated brine (0.5 dm³), and were then dried (MgSO₄), and evaporated under reduced pressure. Crystallization of the gummy residue from hexane gave the title compound (23.56 g, 62.8%) (Found, in material recrystallized from methanol-water: C, 52.89; H, 9.12. C₂₄H₄₈O₉Si₂·0.5 H₂O requires C, 52.81; H 9.05%); mp 133–134 °C; $R_f 0.49$ (ethyl acetate); δ_H (CDCl₃) 1.04 (28 H, m), 1.82 (1 H, m), 1.96 (3 H, m), 2.54 (1 H, d, J 0.8), 2.71 (1 H, d, J 3.6), 3.36 (3 H, s), 3.46 (3 H, m), 3.69 (4 H, m), 3.81 (1 H, m), 3.91 (2 H, m), 4.40 (1 H, t, J 2.4); $\delta_{\rm C}$ (CDCl₃) 12.13, 12.19, 12.82, 17.22, 17.26, 17.29, 17.35, 17.36, 17.39, 17.40, 33.26, 34.53, 49.66, 64.72, 64.96, 70.95, 71.66, 72.28, 74.60, 76.37, 78.30, 99.81.

6-*O*-{2,7-Dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-4,5-*O*-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-*myo*-inositol 18

A mixture of 2-*O*-(4-methoxytetrahydropyran-4-yl)-4,5-*O*-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-*myo*-inositol **17** (10.62 g, 19.8 mmol), 2,7-dibromo-9-chloro-9-[3-(trifluoro-methyl)phenyl]xanthene²⁶ (DtpxCl) (20.49 g, 39.5 mmol) and pyridine (10 cm³) were stirred at room temperature for 20 min. Acetonitrile (70 cm³) was then added over a period of 1 h to the stirred reactants. Water (1.4 cm³) and triethylamine (6.0 cm³, 43 mmol) were added and the products were evaporated under reduced pressure. A solution of the residue in ethyl acetate (500 cm³) was back-extracted with ethyl acetate (100 cm³). The

 $[\]ddagger$ 4-Methoxy-3,6-dihydro-2*H*-pyran **14** (\approx 70%) was recovered by distillation of the filtrate.

combined organic layers were washed with brine (100 cm³), dried (MgSO₄), and evaporated under reduced pressure. Acetonitrile (≈250 cm³) was added and the crystalline precipitate (3.14 g), believed to be the 1-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl} derivative, was collected by filtration. The filtrate was evaporated and the residue was chromatographed on silica gel. The column was eluted with hexane-diethyl ether (100:0 to 0:100 v/v). More of the putative 1-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]isomeric xanthen-9-yl} derivative, which had $R_f 0.55$ [dichloromethanemethanol (98:2 v/v)], was eluted first. The title compound (14.88 g, 73%) [Found: $(M - H)^-$ (FAB), 1017.1849. ¹²C₄₄¹H₅₆⁷⁹Br⁸¹Br¹⁹F₃¹⁶O₁₀²⁸Si₂ requires *m/z*, 1017.1710] had $R_{\rm f}$ 0.25 [dichloromethane-methanol (98:2 v/v)]; $\delta_{\rm H}$ (CDCl₃) 1.12 (28 H, m), 1.48 (1 H, m), 1.76 (3 H, m), 2.58 (1 H, br), 2.81 (1 H, d, J 8.0), 2.91 (3 H, s), 3.18 (1 H, dt, J 2.6 and 8.3), 3.44 (1 H, d, J 8.6), 3.60 (3 H, m), 3.78 (2 H, m), 3.89 (1 H, t, J 8.6), 3.97 (1 H, t, J 8.4), 4.08 (1 H, t, J 2.3), 7.02 (1 H, d, J 8.7), 7.09 (1 H, d, J 8.8), 7.37 (7 H, m), 7.80 (1 H, d, J 8.4); $\delta_{\rm C}$ (CDCl₃) included the following signals: 12.46, 12.60, 12.68, 13.45, 17.28, 17.34, 17.36, 17.41, 17.54, 17.67, 17.78, 18.02, 32.98, 34.36, 48.97, 64.58, 64.79, 71.33, 71.51, 72.18, 75.94, 76.56, 77.83, 79.00, 99.53.

6-*O*-{2,7-Dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-3-*O*-(-)-menthoxyacetyl-2-*O*-(4-methoxytetrahydropyran-4-yl)-4,5-*O*-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-D- and -L-*myo* inositol 19 and 20

Dry pyridine (80 cm³) was added slowly to a stirred mixture of (-)-MacCl (6.30 g, 27.1 mmol) and freshly sublimed 1*H*-tetrazole (1.98 g, 28.3 mmol) in dry acetonitrile (15 cm³) at room temperature. The resulting solution was added rapidly to a mixture of 6-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-O-(4-methoxytetrahydropyran-4-yl)-4,5-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-myo-inositol 18 (14.62 g, 14.3 mmol) and DMAP (4.94 g, 40.4 mmol). After 45 min, when TLC [hexane-diethyl ether (1:1 v/v)] revealed that the proportions of starting material $(R_f 0.12)$ and the putative bis[(-)-menthoxyacetate] ($R_{\rm f}$ 0.64) were approximately equal, water (1 cm³) was added. The products were evaporated under reduced pressure and the residue was re-evaporated with toluene $(3 \times 50 \text{ cm}^3)$. Ethyl acetate (250 cm^3) was added, and after filtration the solution was washed with saturated aq. sodium hydrogen carbonate $(2 \times 150 \text{ cm}^3)$. The combined aqueous layers were back-extracted with ethyl acetate (100 cm³). Finally, the combined organic layers were washed successively with water $(2 \times 100 \text{ cm}^3)$ and brine (100 cm^3) , and were then dried (MgSO₄), and evaporated under reduced pressure.

When the residual glass was dissolved in acetonitrile, colourless crystals of $6-O-\{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]$ $xanthen-9-yl}-3-O-(-)-menthoxyacetyl-2-O-(4-methoxytetra$ hydropyran-4-yl)-4,5-O-(1,1,3,3-tetraisopropyldisiloxane-1,3diyl)-L-myo-inositol**20**were obtained (5.96 g, 34.2%) (Found,in material recrystallized from acetonitrile-diethyl ether: C,55.2; H, 6.25, CscH₇₇Br₂F₂O₁₂Si, requires C, 55.35; H, 6.39%);

In matchiai (cc) ytamized from accommune-dicting) effect. C, 55.2; H, 6.25. $C_{56}H_{77}Br_2F_3O_{12}Si_2$ requires C, 55.35; H, 6.39%); mp 217 °C; $[a]_D^{25} - 25.3$ (c 2.00, ethyl acetate); R_f 0.38 [hexane-diethyl ether (1:1 v/v)]: δ_H (CDCl₃) 0.73–1.35 (42 H, m), 1.47 (2 H, m), 1.62 (3 H, m), 1.77 (1 H, m), 1.99 (1 H, m), 2.25 (1 H, m), 2.76 (1 H, d, J 8.5), 2.80 (3 H, s), 3.12 (1 H, dt, J 4.1 and 10.5), 3.23 (1 H, dt, J 2.6 and 8.8), 3.51 (2 H, m), 3.64 (2 H, m), 3.82 (1 H, m), 4.05 (5 H, m), 4.89 (1 H, m), 7.00 (1 H, d, J 8.7), 7.08 (1 H, d, J 8.7), 7.26–7.45 (7 H, m), 7.81 (1 H, d, J 7.5); δ_C (CDCl₃) included the following signals: 12.51, 12.59, 12.67, 13.40, 16.10, 17.23, 17.27, 17.43, 17.69, 17.79, 18.02, 20.98, 22.25, 23.14, 25.31, 31.47, 33.30, 34.27, 34.33, 39.65, 48.05, 48.75, 64.49, 64.80, 65.27, 70.68, 71.76, 72.08, 73.65, 75.78, 78.15, 78.64, 79.74, 99.30.

The mother liquors from the above crystallizations (from both acetonitrile and acetonitrile-diethyl ether) were combined, evaporated under reduced pressure, and the residue was fractionated by chromatography on silica gel. The column was eluted with hexane–ethyl acetate (100:0 to 66:34 v/v). Appropriate fractions, eluted with hexane–ethyl acetate (80:20 to 74:26 v/v), were combined, and evaporated under reduced pressure to give $6-O-\{2,7-\text{dibromo-9-}[3-(\text{trifluoro-methyl})\text{phenyl}]$ xanthen-9-yl}-3-O-(-)-menthoxyacetyl-2-O-(4-methoxytetrahydropyran-4-yl)-4,5-O-(1,1,3,3-tetraisopropyl-disiloxane-1,3-diyl)-D-*myo*-inositol **19** as a colourless foam

disiloxane-1,3-diyl)-D-myo-inositol **19** as a colourless foam (4.47 g, 25.6%); $R_{\rm f}$ 0.45 [hexane-diethyl ether (1:1 v/v)]; $[a]_{\rm D}^{25}$ -16.2 (c 2.00, ethyl acetate); $\delta_{\rm H}$ (CDCl₃) 0.74–1.39 (42 H, m), 1.48 (2 H, m), 1.63 (3 H, m), 1.74 (1 H, m), 2.01 (1 H, m), 2.23 (1 H, m), 2.70 (1 H, d, J 8.5), 2.80 (3 H, s), 3.10 (1 H, dt, J 4.2 and 10.6), 3.22 (1 H, dt, J 2.6 and 8.8), 3.50 (2 H, m), 3.63 (2 H, m), 3.81 (1 H, m), 4.05 (5 H, m), 4.84 (1 H, m), 7.00 (1 H, d, J 8.7), 7.08 (1 H, d, J 8.7), 7.30–7.45 (7 H, m), 7.80 (1 H, d, J 7.6); $\delta_{\rm C}$ (CDCl₃) included the following signals: 12.52, 12.63, 12.69, 13.43, 14.13, 16.13, 17.21, 17.26, 17.28, 17.43, 17.70, 17.81, 18.03, 20.97, 22.23, 23.16, 25.32, 26.92, 31.55, 33.41, 34.29, 34.34, 39.83, 48.04, 48.68, 64.50, 64.80, 65.65, 70.68, 71.70, 72.36, 73.58, 75.78, 78.23, 78.67, 80.25, 99.31.

Later fractions, which were eluted from the column with hexane–ethyl acetate (73:27 to 66:34 v/v), were concentrated under reduced pressure. The residue was crystallized from acetonitrile to give more L-*myo*-inositol-derived diastereo-isomer **20** [1.33 g; total yield of this diastereoisomer: 7.29 g (41.7%)]. The mother liquor was concentrated and rechromatographed to give more D-*myo*-inositol-derived diastereoisomer **19** [2.62 g; total yield of this diastereoisomer, 7.09 g (40.5%)].

6-*O*-{2,7-Dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-D-*myo*-inositol 21

A solution of 6-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-3-O-(-)-menthoxyacetyl-2-O-(4-methoxytetrahydropyran-4-yl)-4,5-O-(1,1,3,3-tetraisopropyldisiloxane-1,3diyl)-D-myo-inositol 19 (7.09 g, 5.8 mmol) in 8 mol dm^{-3} ethanolic methylamine (40 cm3) was stirred at room temperature for 90 min and was then evaporated under reduced pressure. The residue was dissolved in a 1.0 mol dm⁻³ solution of TEAF in acetonitrile (25 cm³) at room temperature. After 30 min, the products were concentrated under reduced pressure and the residue was fractionated between ethyl acetate (300 cm³) and half-saturated aq. sodium hydrogen carbonate (200 cm³). The organic layer was separated, washed successively with water $(2 \times 100 \text{ cm}^3)$ and brine (50 cm³), dried (MgSO₄), and evaporated under reduced pressure. The residue was fractionated by chromatography on silica gel (200 g). The column was eluted first with dichloromethane-acetonitrile (100:0 to 50:50 v/v). Subsequent elution with dichloromethane-methanol (96:4 to 89:11 v/v) and evaporation of the appropriate fractions gave the title compound (4.33 g, $\approx 95\%$) as a colourless foam; $R_f 0.42$ [dichloromethane–ethanol (9:1 v/v)]; $\delta_{\rm H}$ [(CD₃)₂SO] 1.55–1.75 (4 H, m), 3.05 (3 H, s), 3.10 (1 H, m), 3.30 (5 H, m), 3.50 (1 H, t, J 7.5), 3.56 (2 H, m), 3.99 (1 H, m), 4.19 (1 H, d, J 4.7), 4.27 (1 H, d, J 5.7), 4.52 (1 H, d, J 4.1), 4.70 (1 H, m), 7.08 (1 H, d, J 2.3), 7.16 (1 H, d, J 2.4), 7.20 (2 H, t, J 9.0), 7.41 (1 H, d, J 8.0), 7.48 (3 H, m), 7.60 (1 H, d, J 7.6), 8.07 (1 H, br s); $\delta_{\rm C}$ [(CD₃)₂SO] included the following signals: 34.32, 34.44, 48.37, 64.27, 71.14, 73.45, 73.67, 75.12, 76.76, 98.33.

6-*O*-{2,7-Dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-L-*myo*-inositol 22

A suspension of crystalline 6-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-3-O-(-)-menthoxyacetyl-2-O-(4methoxytetrahydropyran-4-yl)-4,5-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-L-*myo*-inositol **20** (1.215 g, 1.0 mmol) in a 1.0 mol dm⁻³ solution of TEAF in acetonitrile (4 cm³) was heated, under gentle reflux, for *ca*. 2 min until a homogeneous solution was obtained. The cooled products were evaporated under reduced pressure and the residue was dissolved in ethyl acetate (100 cm³). The solution was washed first with water (2 × 30 cm³) and then with an aqueous solution containing both sodium chloride and sodium hydrogen carbonate (30 cm³). The dried (MgSO₄) organic layer was concentrated under reduced pressure and the residue was dissolved in absolute ethanol (5 cm³) at room temperature. Ethanolic methylamine (8.0 mol dm⁻³; 5 cm³) was then added. After 40 min, the products were evaporated under reduced pressure and fractionated by chromatography on silica gel as in the above preparation of the enantiomeric inositol derivative **21**. The title compound **22** (0.646 g, ≈83%) was obtained as a colourless glass; its ¹H and ¹³C NMR spectra were identical with the corresponding spectra of its enantiomer.

1,4,5,6-Tetra-O-benzyl-D-myo-inositol 26

Chloroacetic acid (0.37 g, 3.9 mmol) was added to a stirred solution of 6-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-3-O-(-)-menthoxyacetyl-2-O-(4-methoxytetrahydropyran-4-yl)-4,5-O-(1,1,3,3-tetraisopropyldisiloxane-1,3diyl)-D-myo-inositol 19 (0.243 g, 0.02 mmol) and pyrrole (0.20 cm³, 2.9 mmol) in dry dichloromethane (10 cm³) at room temperature. After 15 min, triethylamine (2.0 cm³) and dichloromethane (50 cm³) were added and the products were washed with saturated aq. sodium hydrogen carbonate (100 cm³). The aqueous layer was separated, and back-extracted with dichloromethane (50 cm^3). The combined organic layers were dried (MgSO₄), and evaporated under reduced pressure. The residue was fractionated by chromatography on silica gel (10 g). The column was eluted with hexane-ethyl acetate (100:0 to 50:50 v/v): the appropriate fractions, which were eluted with hexane-ethyl acetate (70:30 to 60:40 v/v), were combined, and evaporated under reduced pressure to give a colourless glass (0.132 g); $R_f 0.26$ [hexane-ethyl acetate (7:3 v/v)].

This material (0.132 g) and imidazole (0.049 g, 0.72 mmol) were dissolved in dry DMF (1.0 cm³) at room temperature and 1,3-dichloro-1,1,3,3-tetraisopropyldisiloxane **16** (0.115 cm³, 0.36 mmol) was added to the well stirred solution. The reactants were heated at 80 °C. After 7 h, the products were cooled and ethyl acetate (100 cm³) was added. The resulting solution was washed successively with half-saturated aq. sodium hydrogen carbonate (2×100 cm³), water (3×100 cm³) and brine (50 cm³). The combined aqueous layers were back-extracted with ethyl acetate (50 cm³). The combined organic extracts were dried (MgSO₄), and concentrated under reduced pressure. Crystallization from acetonitrile gave colourless crystals of compound **23** (0.119 g), mp 160–162 °C; R_f 0.46 [hexane–diethyl ether (4:1 v/v)].

This material (0.423 g, obtained from more than one experiment) and PTSA monohydrate (≈ 0.005 g) were dissolved in dichloromethane–methanol (1:1 v/v; 5 cm³) at room temperature. After 2 h, triethylamine (2 cm³) was added and the products were evaporated under reduced pressure. The residue was dissolved in ethanolic methylamine (8 mol dm⁻³, 10 cm³). After 45 min, the products were concentrated under reduced pressure and the residue was recrystallized from acetonitrile to give colourless crystals (0.155 g); R_f 0.26 [hexane–diethyl ether (4:1 v/v)]. A second crop (0.098 g) was obtained following chromatography of the mother liquors.

The product (0.202 g) and PTSA monohydrate (≈ 0.005 g) were dissolved in 1,1-dimethoxycyclopentane (1.0 cm³) at room temperature. After 1 h, triethylamine (0.2 cm³) was added and the products were concentrated under reduced pressure. The residue was dissolved in hexane (30 cm³) and silica gel (2 g) was added to the stirred solution. After 1 min, the products were filtered and the residue was washed with hexane–diethyl ether (95:5 v/v; 100 cm³). The filtrate and washings were evaporated under reduced pressure and the residue (compound **25**) was

dissolved in ethanol (2 cm³). An acetonitrile solution of TEAF (1 mol dm⁻³, 80 mm³) and aq. tetraethylammonium hydroxide (20% w/v; 49 mm³) were added and the reactants were heated, under gentle reflux, for 16 h. The products were neutralized by bubbling in carbon dioxide gas and were then concentrated. The residual gum was triturated with diethyl ether $(3 \times 10 \text{ cm}^3)$ and the supernatant liquid was discarded. The remaining material was evaporated from acetonitrile solution $(3 \times 5 \text{ cm}^3)$ and was then dissolved in DMF (2 cm³). This solution was added carefully to sodium hydride (60% dispersion in mineral oil; 0.389 g, 9.7 mmol) at 0 °C (ice-water-bath) and benzyl chloride (0.55 cm³, 4.8 mmol) was added dropwise over a period of 1 min to the stirred resulting mixture. After 2.5 h, ethanol was added dropwise until the effervescence ceased. Ethyl acetate (100 cm³) was then added. The resulting mixture was washed successively with half-saturated aq. sodium hydrogen carbonate $(2 \times 50 \text{ cm}^3)$, water $(3 \times 50 \text{ cm}^3)$ and brine (50 cm^3) ; it was then dried (MgSO₄), and evaporated under reduced pressure. The residue was fractionated by silica gel chromatography. The column was eluted with hexane-diethyl ether (100:0 to 80:20 v/v): the appropriate fractions, which were eluted with hexane-diethyl ether (96:4 to 80:20 v/v), were evaporated under reduced pressure. The residue was dissolved in dichloromethane-methanol (1:1 v/v; 6 cm³) at room temperature. Water (0.5 cm³) and PTSA monohydrate (0.02 g) were added. After 2 h, triethylamine (0.2 cm³) was added and the products were concentrated under reduced pressure. The residue was dissolved in ethyl acetate (100 cm³). The resulting solution was washed successively with half-saturated aq. sodium hydrogen carbonate $(2 \times 50 \text{ cm}^3)$ and brine; it was then dried (MgSO₄), and evaporated under reduced pressure. Crystallization of the residue from cyclohexane gave the title compound (0.132 g, 19% overall yield for the 8 steps) (Found: C, 75.35; H, 6.6. Calc. for C₃₄H₃₆O₂: C, 75.53; H, 6.71%), mp 144–145 °C (lit.,²⁷ 142.5 °C); R_f 0.08 [hexane–diethyl ether (1:1 v/v)]; $[a]_D^{20}$ +22.7 (c 1.18, chloroform) [lit.,²⁷ +23.4 (c 0.22, chloroform)]; $\delta_{\rm H}$ (CDCl₃) 2.45 (1 H, d, J 4.3), 2.53 (1 H, br s), 3.48 (3 H, m), 3.84 (1 H, t, J 8.5), 3.97 (1 H, t, J 9.5), 4.21 (1 H, m), 4.72 (3 H, m), 4.84 (2 H, dd, J 3.4 and 10.7), 4.94 (3 H, m), 7.31 (20 H, m).

1,4,5,6-Tetra-O-benzyl-L-myo-inositol 27

In the same way, 6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-3-*O*-menthoxyacetyl-2-*O*-(4-methoxytetrahydropyran-4-yl)-4,5-*O*-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-L-*myo*-inositol **20** was converted into 1,4,5,6-tetra-*O*-benzyl-L-*myo*-inositol **27** in 24% overall yield for the 8 corresponding steps. The title compound (Found: C, 75.0; H, 6.7%) had mp 144–145 °C, $[a]_D^{20} - 21.7$ (*c* 1.05, chloroform); its ¹H NMR spectrum was identical with that of its D-enantiomer **26** (see above).

3,4,5-Tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-*D-myo*-inositol 29

CpacCl (2.81 cm³, 18.0 mmol) was added to a stirred suspension of 1*H*-tetrazole (1.33 g, 19.0 mmol) in dry acetonitrile (15 cm³) at room temperature. Pyridine (7.5 cm³) was then added slowly. The resulting yellow–orange solution was added to $6-O-\{2,7-\text{dibromo-9-}[3-(\text{trifluoromethyl})\text{phenyl}]\text{xanthen-9-yl}-2-O-(4-methoxytetrahydropyran-4-yl)-D-$ *myo*-inositol**21**(3.12 g, 4.0 mmol) and the reactants were stirred at room temperature for 1 h. DMAP (2.66 g, 21.8 mmol) was then added and the reactants were stirred at room temperature for a further period of 35 min. Water (0.33 cm³) was added and the products were concentrated under reduced pressure. The residue was suspended in ethyl acetate (200 cm³) and the mixture was filtered through a bed of Celite. The filtrate was washed first with saturated aq. sodium hydrogen carbonate

 $(2 \times 100 \text{ cm}^3)$ and then with brine-0.20 mol dm⁻³ sodium acetate buffer (pH 5; 100 cm³). The dried (MgSO₄) organic layer was evaporated under reduced pressure and the residue was fractionated by chromatography on silica gel. The column was washed with hexane-dichloromethane-triethylamine (0: 99.5:0.5 to 49.5:50:0.5 v/v) and was then eluted with hexanedichloromethane-triethylamine-acetonitrile (49.5:50:0.5:0.0 to 44.5:45:0.5:10 v/v). Early fractions, which were eluted with a solvent mixture containing 1–5% acetonitrile, were combined, and evaporated under reduced pressure to give a product (1.99 g) with R_f 0.48 [diethyl ether-hexane (4:1 v/v)] which was believed to be the tetrakis(4-chlorophenoxy)acetyl] derivative 30. Later fractions, which were eluted with a solvent mixture containing 6-9% acetonitrile, were combined and evaporated under reduced pressure to give the title compound 29 (2.10 g, $\approx 40\%$) as a pale yellow glass, $R_f 0.34$ [diethyl ether-hexane (4:1 v/v)]; δ_H (CDCl₃) 1.54 (2 H, m), 1.75 (2 H, m), 2.53 (1 H, d, J 7.5), 3.08 (3 H, s), 3.52 (3 H, m), 3.70 (2 H, m), 3.78 (1 H, t, J 8.1), 4.37 (5 H, m), 4.47 (2 H, s), 4.99 (1 H, dd, J 2.6 and 9.3), 5.29 (1 H, t, J 8.3), 5.39 (1 H, t, J 8.9), 6.74 (6 H, m), 7.01 (1 H, d, J 2.4), 7.15 (9 H, m), 7.39 (4 H, m), 7.52 (1 H, d, J 7.6), 7.70 (1 H, br s); $\delta_{\rm C}$ (CDCl₃) included the following signals: 33.59, 34.35, 49.39, 64.67, 64.90, 65.28, 69.10, 71.20, 71.54, 71.59, 72.95, 74.85, 100.10.

The above putative tetrakis[(4-chlorophenoxy)acetyl] derivative 30 (3.77 g, obtained from two separate experiments) was dissolved in ethanol (50 cm³) at room temperature and ethanolic methylamine (8 mol dm⁻³; 50 cm³) was added to the stirred solution. After 90 min, the products were concentrated under reduced pressure and fractionated by chromatography on silica gel. The column was first washed with dichloromethaneacetonitrile (100:0 to 50:50 v/v). The appropriate fractions, which were eluted with dichloromethane-methanol (96:4 to 89:11 v/v), were combined, and evaporated under reduced pressure to give 6-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-O-(4-methoxytetrahydropyran-4-yl)-Dmyo-inositol 21 (1.53 g), identical (TLC, ¹H and ¹³C NMR) with the starting material 21 used in the preparation of the above 3,4,5-tris-O-[(4-chlorophenoxy)acetyl] derivative 29. Thus the yield of the latter compound 29 can be estimated to be $\approx 55\%$, based on consumed starting material 21.

3,4,5-Tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-L-*myo*-inositol

 $6-O-\{2,7-\text{Dibromo-9-}[3-(trifluoromethyl)phenyl]xanthen-9-yl\}-2-O-(4-methoxytetrahydropyran-4-yl)-L-$ *myo*-inositol**22**(3.11 g, 4.0 mmol) was allowed to react with 1*H*-tetrazole (1.33 g, 19.0 mmol), CpacCl (2.81 cm³, 18.0 mmol) and DMAP (2.66 g, 21.8 mmol) in pyridine (7.5 cm³) and acetonitrile (15 cm³) as in the above preparation of the corresponding D-enantiomer**29**. The products were worked up and fractionated to give the title compound (2.62 g, 51%; identical TLC, ¹H and ¹³C NMR spectra with the above D-enantiomer**29**) and the corresponding putative tetrakis-*O*-[(4-chlorophenoxy)acetyl] derivative**30**.

Di(2-cyanoethyl) phosphorochloridite 31

A solution of triethylamine (84.5 cm^3 , 0.61 mol) in 3-hydroxypropionitrile (40.3 cm^3 , 0.59 mol) was added dropwise over a period of 20 min to a stirred solution of chlorotrimethylsilane (76.6 cm^3 , 0.60 mol) in dry diethyl ether (500 cm^3) at 0 °C (ice– water-bath). After a further period of 2 h, the products were filtered. The filtrate was concentrated under reduced pressure and then distilled to give 3-(trimethylsilyloxy)propionitrile (80.92 g) as a colourless liquid, bp 78–80 °C/21 mmHg.

The latter compound (8.63 g, ≈ 60 mmol) was dissolved in acetonitrile (30 cm³) at room temperature and phosphorus trichloride (2.39 cm³, 27.4 mmol) was added to the stirred solution. After 2 days, the products were concentrated under reduced pressure (water-pump, followed by oil-pump) to give a colourless oil (6.18 g). This material was estimated by ³¹P NMR spectroscopy { δ_P (CDCl₃) 139.1 [$\approx 20\%$, assigned to the phosphorus resonance of tri(2-cyanoethyl) phosphite], 165.7 ($\approx 75\%$, assigned to the phosphorus resonance of di(2-cyanoethyl) phosphorochloridite], 179.5 ($\approx 5\%$, assigned to the phosphorus resonance of 2-cyanoethyl phosphorodichloridite)} to contain ≈ 75 mol% of the title compound. This material was used without further purification.

3,4,5-Tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-D-*myo*-inositol 1-phosphate di(2-cyanoethyl) ester 33

A solution of the above di(2-cyanoethyl) phosphorochloridite reagent 31 (\approx 75 mol%; 1.45 g) in dry acetonitrile (6 cm³) was added to a stirred solution of 3,4,5-tris-O-[(4-chlorophenoxy)acetyl]-6-O-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-O-(4-methoxytetrahydropyran-4-yl)-D-myoinositol 29 (3.842 g, 3.0 mmol) and 3-nitro-1,2,4-1H-triazole **32** (0.820 g, 7.2 mmol) in dry pyridine (24 cm³) at room temperature. After 1 h, water (0.216 cm³, 12 mmol) was added. The products were concentrated under reduced pressure and the residue was re-dissolved in acetonitrile (20 cm³). tert-Butyl hydroperoxide (70%; 1.64 cm³, 12 mmol) was added to the stirred solution. After 90 min, the products were concentrated to ≈one-third volume under reduced pressure and the residue was dissolved in ethyl acetate (200 cm³). The solution was washed with saturated aq. sodium hydrogen carbonate (2 \times 100 cm³) and the combined aqueous washings were extracted with ethyl acetate (100 cm³). The combined organic layers were washed successively with sodium acetate buffer (pH 5; 0.1 mol dm^{-3} ; 100 cm^{3}) and brine (50 cm³), and were then dried (MgSO₄), and evaporated under reduced pressure. The residue was fractionated by chromatography on silica gel. The column was washed with hexane–dichloromethane (1:1 v/v) and was then eluted with hexane-dichloromethane-ethanol (50: 50:0 to 46:46:8 v/v); the appropriate fractions, which were eluted with solvent mixtures containing 5-7% ethanol, were combined, and evaporated under reduced pressure to give the title compound as an off-white glass (3.765 g, $\approx 85\%$); $[a]_D^{23} + 10.7$ (c 4.01, toluene); $R_{\rm f}$ 0.57 [dichloromethane–ethanol (9:1 v/v)]; Found (M - H - acrylonitrile) (FAB) 1414. ${}^{12}C_{59}{}^{1}H_{49}{}^{79}Br^{81}Br^{35}Cl_{3}$ - ${}^{19}F_{3}{}^{14}N^{16}O_{18}{}^{31}P$ requires m/z, 1414; δ_{H} (CDCl₃) 1.60–1.80 (4 H, m), 2.69 (2 H, t, J 6.2), 2.73 (2 H, m), 3.19 (3 H, s), 3.50 (2 H, m), 3.79 (2 H, m), 4.00–4.32 (7 H, m), 4.4–4.55 (5 H, m), 4.82 (1 H, s), 4.96 (1 H, m), 5.13 (1 H, m), 5.26 (1 H, t, J 7.2), 6.74 (6 H, m), 7.07 (2 H, m), 7.20 (8 H, m), 7.40 (3 H, m), 7.53 $(2 \text{ H}, \text{m}), 7.80 (1 \text{ H}, \text{br}); \delta_{P} (\text{CDCl}_{3}) - 1.6.$

3,4,5-Tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-L-*myo*-inositol 1-phosphate di(2-cyanoethyl) ester 39

3,4,5-Tris-O-[(4-chlorophenoxy)acetyl]-6-O-{2,7-dibromo-9-

[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-O-(4-methoxytetrahydropyran-4-yl)-L-*myo*-inositol **29** (2.615 g, 2.04 mmol), the above di(2-cyanoethyl) phosphorochloridite reagent **31** (\approx 75 mol%; 0.98 g) and 3-nitro-1,2,4-1*H*-triazole **32** (0.562 g, 4.9 mmol) were allowed to react together in dry acetonitrile (4 cm³) and pyridine (12 cm³) solution as in the above preparation of the D-enantiomer **33**. After 80 min, water (0.147 cm³, 8.2 mmol) was added. The products were then treated with *tert*-butyl hydroperoxide (70%; 1.14 cm³, \approx 8.3 mmol) and worked up and chromatographed as above to give the title compound as a colourless glass (2.312 g, \approx 77%); [a]_D²⁰ –10.8 (*c* 3.99, toluene); R_f 0.57 [dichloromethane–ethanol (9:1 v/v)]; its ¹H and ³¹P NMR spectra (CDCl₃) were identical with those of the above D-enantiomer.

Product of the coupling reaction between 3,4,5-tris-O-[(4-chlorophenoxy)acetyl]-6-O-{2,7-dibromo-9-[(3-trifluoromethyl)phenyl]xanthen-9-yl}-O-(4-methoxytetrahydropyran-4-yl)-D-*myo*-inositol 1-phosphate (2-cyanoethyl) ester triethylammonium salt 34 and 2-O-arachidonoyl-1-O-stearoyl-*sn*-glycerol 6; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$

A solution of 3,4,5-tris-O-[(4-chlorophenoxy)acetyl]-6-O-{2,7dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-O-(4methoxytetrahydropyran-4-yl)-D-myo-inositol 1-phosphate di-(2-cyanoethyl) ester 33 (1.76 g, 1.20 mmol) in triethylamineacetonitrile (1:2 v/v; 12 cm³) was stirred at room temperature. After 16 h, the products were concentrated under reduced pressure and the residue was dissolved in dry acetonitrile (10 cm³), and 4-methoxypyridine 1-oxide³⁹ 36 (1.50 g, 12.0 mmol) and 2-O-arachidonoyl-1-O-stearoyl-sn-glycerol 6; $R^1 = C_{17}H_{35}$, $R^2 = C_{17}H_{31}$ (1.52 g, 2.4 mmol) were added. The resulting solution was evaporated under reduced pressure. The residue was re-dissolved in dry acetonitrile (10 cm³) and the solution was re-evaporated. After this process had been repeated once more, the residue was redissolved in dry acetonitrile (4 cm³). A solution of mesitylene-2-sulfonyl chloride **35** (1.31 g, 6.0 mmol) in dry pyridine (8 cm³) was added dropwise over a period of 30 min to the stirred solution at room temperature. After a further period of 30 min, water (0.22 cm³) was added and the products were concentrated under reduced pressure. The residue was dissolved in ethyl acetate (200 cm³) and the resulting solution was washed with saturated aq. sodium hydrogen carbonate $(2 \times 100 \text{ cm}^3)$. The aqueous washings were back-extracted with ethyl acetate (100 cm³). The combined organic layers were washed successively with 0.1 mol dm^{-3} sodium acetate buffer (pH 5; 2 × 100 cm³) and brine (50 cm³) and were then dried (MgSO₄), and evaporated under reduced pressure. The residue was fractionated by chromatography on silica gel. The column was eluted with hexane-dichloromethane-ethanol (50:50:0 to 48.5:48.5:3 v/v); the appropriate fractions, which were eluted with solvent mixtures containing 1.5-2.5% ethanol, were evaporated under reduced pressure to give the product 37; $R^1 = C_{17}H_{35}$, $R^2 =$ $C_{19}H_{31}$ (1.75 g, 71%); $R_f 0.44$ [dichloromethane-ethanol (95:5 v/v)]: $[a]_{D}^{22}$ + 3.3 (c 3.84, ethyl acetate); δ_{H} (CDCl₃) 0.88 (6 H, m), 1.10-1.45 (34 H, m), 1.58-1.76 (8 H, m), 2.07 (4 H, m), 2.30 (4 H, m), 2.67 (2 H, m), 2.81 (6 H, m), 3.17 (3 H, s), 3.49 (2 H, m), 3.76 (2 H m), 4.05-4.30 (9 H, m), 4.38-4.54 (5 H, m), 4.79 (1 H, m), 5.0–5.42 (12 H, m), 6.72 (6 H, m), 7.0–7.22 (10 H, m), 7.36 (3 H, m), 7.50 (2 H, m), 7.77 (1 H, m); $\delta_{\mathbf{P}}$ (CDCl₃) -0.89, -0.73.

Fully protected 1-*O*-(2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycero-3-phospho)-D-*myo*-inositol 3,4,5-trisphosphate 38; $R^{1} = C_{17}H_{35}, R^{2} = C_{19}H_{31}$

Hydrazine monohydrate (0.189 cm³, 3.9 mmol) was added to a stirred solution of compound 37 (1.746 g, 0.86 mmol) in acetonitrile (8 cm³) at room temperature. After 90 min, acetone (8 cm³) was added. After a further period of 2 min, the products were evaporated under reduced pressure and hexane (100 cm³) was added. The crystalline precipitate was removed by filtration and washed with hexane. The combined filtrate and washings were concentrated under reduced pressure. The residue was fractionated by chromatography on silica gel. The column was eluted with ethyl acetate-hexane (2:8 to 10:0 v/v); the appropriate fractions, which were eluted with 100% ethyl acetate, were combined, and evaporated under reduced pressure to give a colourless gum (1.137 g). This material was evaporated with dry acetonitrile $(3 \times 10 \text{ cm}^3)$ and then dissolved in dry pyridine (4.5 cm³). To the resulting solution, which was maintained at room temperature, was added 3-nitro-1,2,41H-triazole 32 (0.768 g, 6.7 mmol), followed by di(2-cyanoethyl) phosphorochloridite reagent 31 (~75 mol%; 1.316 g) in acetonitrile (8 cm³) solution. After 1 h, water (0.10 cm³) was added and the products were evaporated under reduced pressure. The residue was re-dissolved in acetonitrile (10 cm³) at 0 °C (ice-water-bath) and tert-butyl hydroperoxide (70%; 1.08 cm³, \approx 7.8 mmol) was added to the stirred solution. After 5 min, the products were allowed to warm to room temperature and, after 1 h, water was added until the solution became turbid. The products were then applied directly to a column of silanized silica gel. The column was eluted with acetonitrilewater (2:8 to 10:0 v/v): the appropriate fractions, which were eluted with 100% acetonitrile, were combined, and evaporated under reduced pressure. The residue was rechromatographed on a standard silica gel column which was eluted with dichloromethane-propan-2-ol-acetic acid (99.7:0:0.3 to 89.7:10:0.3 v/v): the appropriate fractions, which were eluted with 6–9% propan-2-ol, were combined, and evaporated under reduced pressure to give the title product as a colourless gum (1.027 g, $\approx 57\%$ overall yield for the three steps); $R_{\rm f}$ 0.26 [dichloromethane–ethanol (19:1 v/v)]; $[a]_{D}^{20}$ +25.9 (c 5.1, ethyl acetate); $\delta_{\mathbf{P}}$ (CDCl₃) -3.68, -3.37, -2.01, -1.78, -1.75, -1.72, -1.68.

1-O-(2-O-Arachidonoyl-1-O-stearoyl-*sn*-glycero-3-phospho)-D-*myo*-inositol 3,4,5-trisphosphate [PtdIns(3,4,5)P₃] 2

The above fully protected 1-O-(2-O-arachidonoyl-1-Ostearoyl-sn-glycero-3-phospho-D-myo-inositol 3,4,5-trisphosphate 38; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$ (0.254 g, 0.12 mmol) was coevaporated with dry toluene $(3 \times 5 \text{ cm}^3)$ under reduced pressure and then dissolved in dry acetonitrile (3 cm³). TMG (0.27 cm³, 2.2 mmol) and chlorotrimethylsilane (0.22 cm³, 1.7 mmol) were added and the mixture was stirred at room temperature. After 16 h, the products were evaporated under reduced pressure (oil-pump) and the residue was co-evaporated with dry toluene $(3 \times 5 \text{ cm}^3)$. The residue was triturated with hexane $(3 \times 15 \text{ cm}^3)$ and the resulting mixture was filtered in an atmosphere of argon. The slightly turbid filtrate was concentrated under reduced pressure. The residual colourless glass obtained was dissolved in conc. aq. ammonia (d 0.88)-methanol (5.5:94.5 v/v; 0.86 cm³), and the solution obtained was evaporated under reduced pressure. The colourless residue was dissolved in acetic acid-water (2:1 v/v; 6 cm³) and the resulting solution was stirred at room temperature. After 90 min, the products were evaporated under reduced pressure and the residue was coevaporated with ethanol $(3 \times 10 \text{ cm}^3)$. The powdery residue was triturated first with acetonitrile and then with ethyl acetate. An aqueous solution of this material was passed through an Amberlite IR-120 (plus) cation-exchange resin column (H⁺ form; 20 cm \times 2.5 cm diameter). The acidic eluate was carefully neutralized with methanolic ammonia and the resulting solution was evaporated under reduced pressure. The residue was co-evaporated with dry ethanol to give the ammonium salt of 1-O-(2-O-arachidonoyl-1-O-stearoyl-snglycero-3-phospho)-D-myo-inositol [PtdIns $(3,4,5)P_3$] 2 as an off-white hygroscopic powder (0.146 g, ≈96% yield, assuming that the product is an unhydrated hepta-ammonium salt); δ_H (CD₃OD–D₂O) 0.86 (6 H, m), 1.30 (32 H, m), 1.55 (2 H, m), 1.68 (2 H, m), 2.05 (4 H, m), 2.27–2.47 (4 H, m), 2.78 (6 H, m), 3.90 (1 H, t, J 9.5), 4.0-4.15 (6H, m), 4.23 (1 H, dd, J 8.2 and 11.6), 4.41 (3 H, m), 5.34 (8 H, m); δ_P (CD₃OD–D₂O) 0.02, 1.30, 1.91, 2.22 [Found: (M – H)⁻ (MALDI-TOF) 1125.49. ${}^{12}C_{47}{}^{1}H_{85}{}^{16}O_{22}{}^{31}P_{4}$ requires m/z, 1125.45].

Products of other coupling reactions between protected D- and Lmyo-inositol 1-phosphate (2-cyanoethyl) esters and enantiomeric di-O-acylglycerol derivatives

(a) A fully protected phosphatidylinositol derivative (0.639 g, 69%) was prepared in the same way as the above fully protected

natural diastereoisomer **37**; $R^1 = C_{17}H_{35}$, $R^2 = C_{19}H_{31}$ from 3,4,5-tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[(3-trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetra-hydropyran-4-yl)-D-*myo*-inositol 1-phosphate di(2-cyano-ethyl) ester **33** (0.658 g, 0.45 mmol) and 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8** (0.578 g, 0.90 mmol); it had $[a]_D^{20}$ -0.3 (*c* 4.3, ethyl acetate) and δ_P (CDCl₃) -1.16, -0.63.

(b) A fully protected phosphatidylinositol derivative (0.834 g, 68%) was prepared in the same way as above from 3,4,5-tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-L-*myo*-inositol 1-phosphate di(2-cyanoethyl) ester **39** (0.871 g, 0.59 mmol) and 2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycerol **6** (0.765 g, 1.19 mmol); it had $[a]_{\rm D}^{21}$ +0.7 (*c* 4.6, ethyl acetate) and $\delta_{\rm P}$ (CDCl₃) -1.16, -0.61.

(c) A fully protected phosphatidylinositol derivative (0.692 g, 75%) was prepared in the same way as above from 3,4,5-tris-*O*-[(4-chlorophenoxy)acetyl]-6-*O*-{2,7-dibromo-9-[3-(trifluoromethyl)phenyl]xanthen-9-yl}-2-*O*-(4-methoxytetrahydropyran-4-yl)-L-*myo*-inositol 1-phosphate di(2-cyanoethyl) ester **39** (0.663 g, 0.45 mmol) and 2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycerol **8** (0.580 g, 0.90 mmol); it had $[a]_{20}^{20} - 1.5$ (*c* 3.6, ethyl acetate) and $\delta_{\rm P}$ (CDCl₃) -0.87, -0.71.

Other fully protected 1-*O*-(2-*O*-arachidonoyl-1(or 3)-*O*-stearoylsn-glycero-3-phospho)-D (or L)-myo-inositol 3,4,5-trisphosphates

The diastereoisomers of the fully protected 1-O-(2-O-arachidonoyl-1-O-stearoyl-*sn*-glycero-3-phospho)-D-*myo*-inositol

3,4,5-trisphosphate **38** were prepared from the products of the other coupling reactions between protected D- and L-*myo*-inositol 1-phosphates (2-cyanoethyl) esters and enantiomeric di-O-acylglycerol derivatives described above.

(a) The coupling-reaction product (0.639 g, 0.43 mmol) described above under heading (a) was subjected to the same three-step process [*i.e.*, treatment with (i) hydrazine hydrate, (ii) di(2-cyanoethyl) phosphorochloridite and (iii) *tert*-butyl hydroperoxide] described above in the preparation of the natural diastereoisomer **38** to give fully protected 1-*O*-(2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycero-1-phospho)-D-*myo*-inositol 3,4,5-trisphosphate (0.206 g, 31%); $[a]_{20}^{20}$ +17.8 (*c* 4.1, ethyl acetate); δ_{P} (CDCl₃) -3.65, -3.33, -2.14, -1.80, -1.75, -1.52.

(b) In the same way, the coupling reaction product (0.692 g, 0.47 mmol) described above under heading (b) was converted into fully protected 1-*O*-(2-*O*-arachidonoyl-1-*O*-stearoyl-*sn*-glycero-3-phospho)-L-*myo*-inositol 3,4,5-trisphosphate (0.238 g, 33%); $[a]_{20}^{20}$ -16.4 (*c* 3.5, ethyl acetate); $\delta_{\rm P}$ (CDCl₃) -3.67, -3.35, -2.00, -1.86, -1.80, -1.77, -1.71.

(c) In the same way, the coupling reaction product (0.644 g, 0.44 mmol) described above under heading (c) was converted into fully protected 1-*O*-(2-*O*-arachidonoyl-3-*O*-stearoyl-*sn*-glycero-1-phospho)-L-*myo*-inositol 3,4,5-trisphosphate (0.273 g, 40%); $[a]_{D}^{20}$ -26.8 (c 3.5, ethyl acetate); $\delta_{\rm P}$ (CDCl₃) -3.68, -3.37, -2.00, -1.81, -1.78, -1.75, -1.73, -1.69.

Other diastereoisomers of 1-*O*-(2-*O*-arachidonoyl-1-*O*-stearoyl*sn*-glycero-3-phospho)-D-*myo*-inositol 3,4,5-trisphosphate [PtdIns(3,4,5)*P*₃] 2

The three above fully protected diastereoisomers of 1-O-(2-O-arachidonoyl-1-O-stearoyl-sn-glycero-3-phospho)-D-myo-

inositol 3,4,5-trisphosphate were unblocked by the same threestep process [*i.e.*, treatment with (i) TMG and chlorotrimethylsilane, (ii) methanolic ammonia and (iii) acetic acid-water (2:1 v/v) used in the unblocking of fully protected PtdIns- $(3,4,5)P_3$ **38** itself].

(a) 1-O-(2-O-Arachidonoyl-3-O-stearoyl-*sn*-glycero-1-phospho)-D-*myo*-inositol 3,4,5-trisphosphate **42** (0.077 g, \approx 78% yield, assuming that the product is an unhydrated hepta-ammonium salt) was thereby obtained from its fully protected precursor (0.206 g, 0.10 mmol); $\delta_{\rm P}$ (CD₃OD–D₂O) 1.60, 2.51, 3.05, 3.33.

(b) 1-O-(2-O-Arachidonoyl-1-O-stearoyl-*sn*-glycero-3-phospho)-L-*myo*-inositol 3,4,5-trisphosphate **41** (0.137 g, $\approx 97\%$ yield, assuming that the product is an unhydrated hepta-ammonium salt) was thereby obtained from its fully protected precursor (0.236 g, 0.11 mmol); $\delta_{\rm P}$ (CD₃OD–D₂O) 4.01, 5.10, 5.68, 5.98.

(c) 1-O-(2-O-Arachidonoyl-3-O-stearoyl-*sn*-glycero-1-phospho)-L-*myo*-inositol 3,4,5-trisphosphate **43** (0.159 g, $\approx 97\%$ yield, assuming that the product is an unhydrated hepta-ammonium salt) was thereby obtained from its fully protected precursor (0.273 g, 0.13 mmol); $\delta_{\rm P}$ (CD₃OD–D₂O) 1.62, 2.83, 3.48, 3.73.

Acknowledgements

We thank the Wellcome Trust for generous financial support; we also thank Dr Len Stephens and Professor Peter Downes for valuable discussions and for providing support for one of us (P. R. J. G.). We also thank Dr Alex Drake for CD spectroscopic measurements.

References

- 1 H. Streb, R. F. Irvine, M. J. Berridge and I. Schulz, *Nature*, 1983, **306**, 67.
- 2 B. A. Hemmings, Science, 1997, 277, 534.
- 3 C. B. Reese and J. G. Ward, Tetrahedron Lett., 1987, 28, 2309.
- 4 B. V. L. Potter and D. Lampe, Angew. Chem., Int. Ed. Engl., 1995, 34, 1932.
- 5 P. R. J. Gaffney and C. B. Reese, *Tetrahedron Lett.*, 1997, **38**, 2593.
- 6 P. R. J. Gaffney and C. B. Reese, *Bioorg. Med. Chem. Lett.*, 1997, 7, 3171.
- 7 A. Toker, M. Meyer, K. K. Reddy, J. R. Falck, S. Aneja, D. J. Burns, L. M. Bellas and L. C. Cantley, J. Biol. Chem., 1994, 269, 32358.
 ⁸ K. K. Baddy, M. Saady, J. P. Falck and C. Whited, J. Org. Cham.
- 8 K. K. Reddy, M. Saady, J. R. Falck and G. Whited, *J. Org. Chem.*, 1995, 60, 3385.
 9 Y. Watanaka, H. Hirafaiii and S. Ozaki. *Tetrahaduan Latt.* 1004 25
- 9 Y. Watanabe, H. Hirofuji and S. Ozaki, *Tetrahedron Lett.*, 1994, 35, 123.
- 10 D.-M. Gou and C.-S. Chen, J. Chem. Soc., Chem. Commun., 1994, 2125.
- 11 J. S. Bruzik and R. J. Kuliak, Tetrahedron Lett., 1995, 36, 2415.
- 12 S. G. Aneja, A. Parra, C. Stoenescu, W. Xia and J. Aneja, *Tetrahedron Lett.*, 1997, 38, 803.
- 13 S. J. A. Grove, A. B. Holmes, G. F. Painter, P. T. Hawkins and L. R. Stephens, *Chem. Commun.*, 1997, 1635.
- 14 Y. Watanabe and M. Nakatomi, Tetrahedron Lett., 1998, 39, 1583.
- 15 Y. Watanabe and M. Nakatomi, Tetrahedron, 1999, 55, 9743.
- 16 M. E. Jung and T. J. Shaw, J. Am. Chem. Soc., 1980, 102, 6304.
- 17 C. E. Burgos, D. E. Ayer and R. A. Johnson, J. Org. Chem., 1987, 52, 4973.
- 18 R. G. Jensen and R. G. Pitas, Adv. Lipid Res., 1976, 14, 213.
- 19 M. Gomberg and L. H. Cone, Justus Liebigs Ann. Chem., 1907, 370, 142.
 20 L. M. Braum, C. Christedenkur, C. B. Beau, and C. Sinderen, 1907, 1907.
- 20 J. M. Brown, C. Christodoulou, C. B. Reese and G. Sindona, J. Chem. Soc., Perkin Trans. 1, 1984, 1785.
- 21 J. B. Chatopadhyaya and C. B. Reese, J. Chem. Soc., Chem. Commun., 1978, 640.
- 22 C. B. Reese, H. T. Serafinowska and G. Zappia, *Tetrahedron Lett.*, 1986, **27**, 2291.
- 23 J. J. Baldwin, M. W. Raab, K. Mensler, B. H. Arison and D. E. McClure, J. Org. Chem., 1978, 43, 4876.
- 24 C. B. Reese, R. Saffhill and J. E. Sulston, J. Am. Chem. Soc., 1967, 89, 3366.
- 25 W. T. Markiewicz, J. Chem. Res., 1979, (S) 24; (M) 0181.
- 26 P. R. J. Gaffney, Liu Changsheng, M. V. Rao, C. B. Reese and J. G. Ward, J. Chem. Soc., Perkin Trans. 1, 1991, 1355.
- 27 R. Aneja and A. Parra, Tetrahedron Lett., 1994, 35, 525.
- 28 A. Hampton, J. C. Fratantoni, P. M. Carroll and S. Wang, J. Am. Chem. Soc., 1965, 87, 5481.
- 29 S. S. Jones and C. B. Reese, J. Am. Chem. Soc., 1979, 101, 7399.
- 30 P. Westerduin, G. H. Veeneman and J. H. van Boom, *Recl. Trav. Chim. Pays-Bas*, 1987, **106**, 601.
- 31 A. Jäger and J. Engels, Tetrahedron Lett., 1984, 25, 1437.
- 32 V. A. Efimov, O. G. Chakhmakhcheva and Y. A. Ovchinnikov, *Nucleic Acids Res.*, 1985, **13**, 3651.

- 33 D. A. Evans, R. G. Gage and J. L. Leighton, J. Org. Chem., 1992, 57, 1964.
- 34 D. R. Alessi, S. R. James, C. P. Downes, A. B. Holmes, P. R. J. Gaffney, C. B. Reese and P. Cohen, *Curr. Biol.*, 1997, 7, 261.
 D. R. Alessi, M. Deak, A. Casamayor, F. B. Cauldwell, N. Morris,
- D. G. Norman, P. Gaffney, C. B. Reese, C. N. McDougall, D. Harbison, A. Ashworth and M. Bownes, Curr. Biol., 1997, 7, 776.
- 36 D. Stokoe, L. R. Stephens, T. Copeland, P. R. J. Gaffney,

C. B. Reese, G. F. Painter, A. B. Holmes, F. McCormick and

- P. T. Hawkins, *Science*, 1997, 277, 567.
 J. Stephens, K. Anderson, D. Stokoe, H. Erdjument-Bromage, F. F. Painter, A. B. Holmes, P. R. J. Gaffney, C. B. Reese, E. M. D. T. Hawkins, P. R. J. Gaffney, C. B. Reese, E. M. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, E. M. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, E. M. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, E. M. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, E. M. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, E. M. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, E. M. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. H. L. T. C. B. Reese, I. D. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. H. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. H. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. J. Caffney, C. B. Reese, F. M. L. T. T. Hawkins, P. R. F. McCormick, P. Tempest, J. Coadwell and P. T. Hawkins, Science, 1998, 279, 710.
- 38 V. Bolitt, C. Mioskowski, D.-S. Shin and J. R. Falck, Tetrahedron Lett., 1988, 29, 4583.
- 39 E. Ochiai and M. Katada, J. Pharm. Soc. Jpn., 1943, 63, 265.